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## **Review Article**

## SEED GERMINATION OF ANNATTO (BIXA ORELLANA L.)- A REVIEW

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# ARTICLE INFO ABSTRACT

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#### Key Words:

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Annatto (*Bixa orellana* L.) is a valuable medicinal plant popularly known as "sinduri" or "lipstick tree". The seed coat of Annatto commercially produce dye, a red orange pigment known as bixin which is used mostly in dairy industry, medicine industry, and textile industry and for animal feed. This species is used in production of phytochemicals like flavonoid, sterols, tannins, bisulphate and essential oil. It has been reported that sinduri is used to treat skin problem, liver disease, hepatitis, prostrate disorder, as diuretic and as antioxidant. *Bixa .orellana* L. is commonly propagated through seeds for large scale cultivation. But the seed coat is characterized by water impermeable testa. So it is very difficult to germinate and it also contains below 40% moisture which are the main cause of slow germination. Germination may be increased by using several pretreatments with water, hormones, growth regulators cowdung, acids etc.

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## **INTRODUCTION**

*Bixa orellana*, commonly used for yielding not toxic synthetic dye recommended by WHO and more than 60% of consumed bio colorant produced from annatto (Vilar *et al.*, 2014). The seeds and pulpy layer of seed coat are orange colour due to the presence of bixin. For the enormous demand of organic colour food supplement sinduri are now purchasing priority for conscious fellows. But this plant showed low germination rate in natural environment by many conventional method. To rescue from this problem many workers worked on it, they tried to give optimum viable condition for germination of *Bixa orellana*.

Annatto seeds soaked six to ten days by wet treatments gave significant increase in germination. Many natural common products, like cow dung were used for germination of the plant, they also noticed positive result. Some plant growth promoter (Indole acetic acid, Gibberelic acid and kinetin) were also used as seed germinating stimulant. It was reported that these plant growth promoter influence the seed germination rate of annatto (Castello *et al.*, 2012). Time dependent experiment based on relative humidity and moisture content showed very slow increasing slope on seed germination of *Bixa orellana* (Goldbach, 1979). There were many other tests on seed germination of annatto based on weight and size of the seeds

showed the mean time of germination increased compare with its weight respectively (Joseph *et al.*, 2010).

#### Moisture on Germination of B.orellana

Yogeesha *et al.*, 2005 reported that the maximum germination was found with high moisture level i.e. 60.9% in freshly harvested seeds and lower moisture contents below 12% induced a dormant state. The germination was also reduced to 6% in 12% moisture content. They also reported that fully-imbibed and sterilized seeds produced maximum germination. Again it was noticed that the seed coat of Annatto is also permeable in nature. So the water uptake by seed coat was steadily progressed and stabilized within 24 h. (Joseph *et al.*, 2010).

Many author reported that the seeds of *B.orellana* showed physiological dormancy (Amaral *et al.*, 2000; Amaral *et al.*, 1995; Eira and Mello, 1997; Goldbach, 1979; Yogeesha *et al.*, 2005). However, physical dormancy sometimes interacts with physiological dormancy and ultimately modifies the seed germination (Black *et al.*, 2006). In this regard studying various aspects of germination is important (Joseph *et al.*, 2010).

If the seeds are allowed to dry below 40% moisture content, it become hard seeded i.e. their seed coat are trends to show

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impermeable to water. Freshly harvested mature seeds with 45% moisture content do not show impermeability.

Seed water content and drying period determines the hard seededness ranged from 10% to over 50%. Unclipped seeds germinate very slowly. 18% of the seeds still remained ungerminated but viable keeping one year in the germinator as compared to subsequent germination test after clipping the seeds (Goldbach, 1979).

#### Seed soaking on germination

Seeds soaked in water for different durations (6, 12, 24 and 48 h) and hormone treatments at different concentrations showed significant response of germination. Water soaking from shorter (6 h) to longer (24 h) durations gave a sharp increase in germination of *B.orellana*. Seed soaked in water for 24 h produce higher seed germination (82%). Prolonged soaking for 48 h however produced less germination (70%). On the other hand insufficient soaking with 6 h significantly decreased the germination rate (Joseph *et al.*, 2010).

Seed drying up to 4.2% moisture content significantly reduced germination. Again decrease is found after two month of storage. Open storage at 30% and 50% relative humidity also diminished the germination rate within one year (Goldbach, 1979). On the other hand sulfuric acid treatment at elevated moisture content (60%) enhanced the germination and reduced the mean germination time (MGT). Results are in conformity with several other workers in hard seededness (Das *et al*, 2017).

#### Pretreatment with acid in germination

Germination of *B.orellana* is an important factor during the life cycle of the plant. In Annatto, different results indicated that all pretreatments reduced the percentage of hard seeds as compared to non treated control seeds. The percentage of hard seed reduced gradually as the increment of pretreatment time.

In Annatto, all pretreatments increased the germination rate markedly. Pretreatment with  $H_2SO_4$  in contrast to the control for 15 mins the seed germinates 50% at 9 days. The germination percentage due to pretreatment was reduced as the pretreatment period decreased from 15 min to 5 sec. Further it was found that sulfuric acid pretreatment kill the seeds or populations. When sulfuric acid was used more than 15 mins (30 mins and 1 hour), severe damage in seed was found which leads to less percentage of germination.

Further hydrochloric acid and acetone also found effective in the germination to a certain extent but not fully. In this case many seeds remained hard and ungerminated (Yogeesha *et al.*, 2005).

#### Cow-Dung Water Treatment on germination

Seeds soaked in autoclaved cow dung water did start germination and showed 80% germination. After eleven days radicles are emerged whereas seeds soaked with warm water. Cow-dung is a rich source of nitrogenous compound which is found effective in breaking dormancy such as thiourea and potassium nitrate (Castello *et al.*, 2012).

### Seed mass on germination

Seed mass showed a significant role in germination capacity of Annatto seeds. Medium sized seeds (0.0275-0.03 g) played greater role for germination in comparison to light or heavy

seeds. In light-weight seeds, the lower rate of germination are found due to poor availability of stored food supply (Fox *et al.*, 1994; Siril *et al.*, 1998) compared to heavier seeds (Castro *et al.*, 2007). Due to the hard seed coat low germination of heavy seeds are found (Khan *et al.*, 1999; Susko and Lovett, 2000; Tungate *et al.*, 2002, Joseph *et al.*, 2010).

#### Gibberelic acid $(GA_3)$ in seed germination of B. orellana

 $GA_3$  acid is an initiator of biochemical activities of seed germination. It was also reported that GA has capacity in breaking dormancy. But it did not break dormancy at all in *B. orellana*. Considering the hormone treatments to  $GA_3$  at 50 ppm exhibited more germination (93%) than those to all other treatments. As such,  $GA_3$  treatment resulted in the remarkable reduction of mean time to germination (MTG) (3.6) when soaked for 24 h.  $GA_3$  treatment is then considered to decrease physiological dormancy of *B. orellana* seeds.

From many previous reports (Gupta, 2003), it was found that the proportion of viable seeds found to be 20% in an annatto seed lot. Further germination was reported ranging from 5-7% (Sharon and D'Souza, 2000). If dormancy breaking plant hormone (GA<sub>3</sub> at 50 ppm) is used then the germination percentage may increase up to 93%. In GA<sub>3</sub> treated seeds, germination was rapid (MTG 3.6) compared to other soaking treatments. A phytohormone like gibberelin regulates various phases of plant development including seed germination. Gibberellins are synthesized *de novo* during germination (Debaujin and Koorneef, 1999) and it causes its effectiveness (Karseen et al, 1989). Gibberellins regulate endosperm degradation by inducing enzyme activity (Groot and Karseen, 1987) and it leads to early germination of seeds. GA3 effectivity may be attributed to hormone action on cell cycle activation (Groot et al., 1988).

At the time of water uptake the tissue surrounding the embryo imposed the mechanical restrain relieved by exogenous gibberellins promote the germination (Toyomasu *et al.*, 1994). Germination of seeds with coat-induced dormancy can be stimulated by GA<sub>3</sub> (Black *et al.*, 2006). GA<sub>3</sub> showed positive effect on germination and has been commercially used in several crops for raising planting materials in nurseries (Calvo *et al.*, 2004; Ayele *et al.*, 2006).

Physical dormancy breaking treatments are recommended on germination studies of Annatto by several workers (Custodio *et al.*, 2003; Gupta, 2003). It was also noticed that an interaction between physiological dormancy and imbibitions operated and could be tackled by 24 h soaking in a 50 ppm GA<sub>3</sub> solution. This treatment evokes germination by triggering synthesis of different enzymes involved in the process. The heavy, medium and lightweight seeds imbibe and achieve saturation within 24 h of water uptake (Joseph *et al.*, 2010).

It may be stated that for Annatto seeds, a practical recommendation is soaking of seeds in GA<sub>3</sub> (50 ppm) for 24 h prior to germination. The light-weight seeds should be eliminated to enhance germination in the seed lot. It indicates the importance of seed quality in maximizing germination (Joseph *et al.*, 2010).

#### IAA on Germination

IAA treatment irrespective of concentration performed poor in accelerating germination of *B. orellana* seeds. However,

soaking seeds in 10 ppm IAA for 24 h exhibited higher germination (73%) in comparison to 50 ppm and 100 ppm concentration of IAA (Joseph *et al.*, 2010). But about 25% of germinated seedlings showed abnormal swelling of the roots and callus formation by IAA treatment. When auxins were applied externally at higher concentrations swelling of roots were found (Sharon and Kishore, 1975, Amaral *et al.*, 1996, Castello *et al.*, 2012)

#### Other Plant Growth Regulators on germination

Seeds soaked with 500 ppm kinetin gave maximum percentage of germination (90%), amongst all the tried PGRs was observed and produced best seedling growth. Cytokinins also showed better effect to stimulate germination and to break dormancy in different plants (Khan, 1971). Kinetin concentration higher than 500 ppm hampered the growth of seedlings. It also caused senescence of apex in some plants, resulting the total damage of seedlings (Castello *et al.*, 2012).

## CONCLUSION

It is known that the germination of *B.orellana* is problematic. Therefore, different methods as well as fresh seed stock may be considered and study of various aspects of germination is important. The present discussion on *B.orellana* was done to elucidate behavior and mechanisms operating in the germination process. In brief, it has been discussed on water uptake capacity of seeds, effectiveness of seed soaking in gibberellic acid (GA<sub>3</sub>) and indole-3-acetic acid (IAA) on germination, the influence of seed mass on germination, and the extent of variability in germination of seeds collected from different sources.

## References

- Ayele, B.T., Ozga, J.A. and Reinecke, D.M. (2006): Regulation of GA biosynthesis genes during germination and young seedling growth of pea (*Pisum sativum* L.). J. *Plant Growth Regul.* 25:219-232.
- Amaral, L.I.V., M.A. Pereira, and A. L. Cortelazzo. (2000): Germination in developing seeds of *Bixa orellana*. Revista Brasileira de Fisiología Vegetal 12:273-285.
- Amaral, L.I.V., M.A. Pereira and A.L. Cortelazzo. (1995): Dormancy break in seeds of *Bixa orellana*. Revista Brasileira de Fisiologia Vegetal 7:151-157.
- Black, M., Bewley, J.D. and Halmer., P. (2006): The encyclopedia of seeds, science, technology and uses. CAB international, U.K.
- Calvo, A.P., Nicolás, C., Lorenzo, O., Nicolás, G. and Rodriguez, D. (2004): Evidence for positive regulation by gibberellins and ethylene of ACC oxidase expression and activity during transition from dormancy to germination in *Fagus sylvatica* L. seeds. *J. Plant Growth Regul.* 23:44-53.
- Castello, M.C., Sharan, M., and Sharon, M. (2012): In Vitro Culture studies of *Bixa orellana* L: IV-In Vitro and In Vivo Trials for Breaking the Dormancy of Seeds of *Bixa orellana*. 2 (1):174-179.
- Castro, J.,J.A. Hodar and J.M. Gomez. (2007): Seed size. P.397-428. In Handbook of Seed Science and Technology. Basra, A.S. (ed.) Scientific Publishers, Jodhpur, India.

- Custodio, C.C., Neto, N.B., Caseiro, R.F., Ikeda, M., de Clavijo, C.C., Artioli, G.P.I. and Medina, M.A.M. (2003): Anatomy and ultrastructure of somatic embryogenesis of Annatto. *Agronomía Tropical Maracay* 53:33-48.
- Das, M., Sharma, M. and Sivan, P. (2017): Seed Germination and Seedling Vigor Index in *Bixa orellana* and *Clitoria ternatea*. Int. J. Pure Appl. Biosci. 5, 15–19.
- Debaujin, I and M. Koorneef. (1999): Gibberellin requirement for Arabidopsis seed germination is determined by both testa characteristics and embryonic abscisic acid. *Plant Physiol*. 122:415-424
- Eira, M.T. and C.M.C. Mello. (1997): *Bixa orellana* L. seed germination and conservation. *Seed Sci. Technol.* 25:373-380.
- Fox, I.E.D., D.R. Barrett and J.E. Srand. (1994): Germination in *Santalum album* L. Recent research in Western Austrlia and a protocol for Indonesia. *Inter. J. Ecol. Environ. Sci.* 20:345-356.
- Goldbach, H. (1979): Germination and storage of *Bixa* orellana seeds. Seed Sci. Technol. 7:399-402.
- Groot, S. P. C. and Karseen, C. M. (1987): Gibberellins regulate seed germination in tomato by endosperm weakening: a study with gibberellins deficient mutants. *Planta* 71:525-531.
- Groot, S. P. C., Rockicka, K.B., Vermeer, E. and Karseen, C.M. (1988): Gibberellin induced hydrolysis of endosperm cell walls in gibberellin deficient tomato seeds prior to radicle protrusion. *Planta* 174: 500-504.
- Gupta, V. (2003): Seed germination and dormancy breaking techniques for indigenous medicinal and aromatic plants. *J. Med. Aromatic Plant Sei*. 25:402-407.
- Joseph, N., Siril, E.A., and Nair, G.M. (2010): Imbibition Duration, Seed Treatment, Seed Mass and Population Influence Germination of Annatto (*Bixa orellana* L.) Seeds. Seed Technol. 32, 37–45.
- Karseen, C.M., Zagorski, S., Kepcznski, J. and Groot, S.P.C. (1989): A key role of endogenous gibberellins in the control of seed germination. *Ann. Bot.* 63:71-80.
- Khan, M.L., Bhuyan, P., Shnakar, U. andTodaria, N.P. (1999): Seed germination and seedling fitness in *Mesua ferrea* L. in relation to fruit size and seed number per fruit. *Acta Oecol.* 20:599-606.
- Sharon, M. and D'Souza, M.C. (2000): In vitro clonal propagation of annatto (*Bixa orellana* L.). *Current Sci.* 78:1532-1535.
- Sharon, M. and Kishore, H. (1975): Note on the effects of GA IAA and kinetin on the germination of true dormant seeds of potato. *Indian J. of Agril. Sci.*, , 45, 490.
- Siril, E.A., U. Dhar and P.P. Dhyani. 1998: Seed germination of Chinese tallow tree (*Sapium sebiferum*). Forest Farm and Community Tree Res. Rep. 3:55-58.
- Susko, D.J. and Lovett, L.D. (2000): Patterns of seed mass variation and their effects on seedling traits in *Alliaria petiolata* (Brassicaceae). *Am. J. Bot.* 87:56-66.
- Tungate, K.D., Susko, D.J. and Rufty, T.W. (2002): Reproduction and offspring competitiveness of *Senna* obtusifolia influenced by nutrient availability. New Phytol. 154:661-669.
- Toyomasu, T., Yamane, H., Murofushi, N. and Inoue, Y. (1994): Effects of exogenously applied Gibberellin and

red light on endogenous levels of abscisic acid in photoblastic lettuce seeds. *Plant and Physciol. Japanese Soc. Plant Physiol.* 35:127-129.

Vilar, D. de A., Vilar, M.S. de A., Moura, T.F.A. de L. e, Raffin, F.N., Oliveira, M.R. de, Franco, C.F. de O., de Athayde-Filho, P.F., Diniz, M. de F.F.M., and Barbosa-Filho, J.M. (2014). Traditional Uses, Chemical Constituents, and Biological Activities of Bixa orellana L.: A Review. *Sci. World J.* 2014, 1–11.

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Yogeesha, H.S., Shivananda, T.N., and Bhanuprakash, K. (2005): Effect of seed maturity, seed moisture and various pre-treatments on seed germination of annatto (*Bixa orellana* L.). *Seed Sci. Technol.* 33, 97–104.