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Research Article

INVITRO TOXICITY RESPONSES OF BLEPHARISMA INTERMEDIUM TO COMMONLY USED INSECTICIDE DELTAMETHRIN

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ABSTRACT

The pesticides are playing a major role in the contamination of the environment. The present investigation was done on physiological changes and deltamethrin responses of the freshwater ciliate *Blepharisma intermedium* exposed to a selective pyrethroid insecticide. The obtained LC50 values of *Blepharisma intermedium* was found to be 57.54ppm respectively. The acute toxicity tests were done by using probit analysis and worked out sublethal concentrations were found to be 10ppm, 13ppm, 16ppm and 19ppm respectively. The contractile vacuole activity was reduced when the organisms were exposed to different concentrations in time dependent manner. The nuclear changes leading to DNA damage was noted using Feulgen fast green technique (Rizzo and Nooden; 1973). The more abnormal forms were found to be 67.2±0.79 percent at higher concentration 19ppm. The concentrations used for nuclear abnormality studies were 10ppm, 13ppm, 16ppm, 19ppm respectively.

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INTRODUCTION

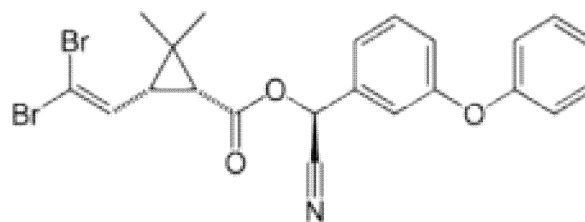
The pesticides are playing a major role in the contamination of soils, aquatic ecosystem, inland and coastal waters and air (Chapalamadugu and Chaudry 1992). Pyrethroids were discovered in 1800's (Rose *et al.*, 2000). Use of pyrethroids is increasing from last three decades (Meacham *et al.*, 2008; Wolansky and Harrill, 2008). Deltamethrin is a type II synthetic pyrethroid (Barahona, 2005; Hossain 2005, Wolansky and Harrill, 2008). Synthetic pyrethroids are highly toxic to zooplanktonic communities (Tidou *et al.* 1992) and to fish (Mestres and Mestres 1992, Mittal *et al.* 1994; Aydin 2004).

Protozoa are eukaryotic and unicellular microorganisms. Freshwater ciliates are being ubiquitous in nature, high reproductive rate, easy to maintain makes them best suited models to screen toxicity effects of various pesticides (Morange, 2006). This organism involves in transformation of nutrients from one trophic level to another trophic level (Hynes, 1960; Carter & Cameron, 1973). Ciliate protozoa act as potential bioindicators of water quality and bio-monitors of ecosystem environment (Masood and Amanchi, 2007, 2008). Tests on the acute toxicity of pollutants on ciliates have revealed that these microorganisms are useful bioindicators for

evaluating the toxicity of water polluted by different pollutants (Madoni, 2011).

MATERIALS AND METHODOLOGY

Deltamethrin (C₂₂H₁₉Br₂NO₃) is an insecticide and its composition 2.8% of Deltamethrin + w/w Emulsifier: 8.00% w/w, Stabilizer (Butylated hydroxytoluene): 1.0% w/w + Solvent (naphtha) 88.20% w/w. manufactured by Bayer Crop science Limited. To carry out the experiments 0.5ml of pesticide solution was added to 4.5ml of culture medium to achieve desired concentration of each pesticide.



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Structure of Deltamethrin

Selected organisms: fresh water ciliate *Blepharisma Intermedium* was used in this study. Pure line stock culture of *Blepharisma intermedium* was supplied from Carolina Biological suppliers, NC, USA. Test organisms were sub cultured in the laboratory and maintained for further studies (Masood and Amanchi, 2008).

Test solutions: The test solutions were prepared according to recommendations given by APHA (2012). Stock solution of deltamethrin 1000ppm was prepared by using distilled water.

Preparation of Culturing Medium: Hay infusion media was used as culture media for growing the ciliates (Kirby, 1950). 6 – 8 grams of hay was added to 1 liter boiling distilled water in a beaker and boiled for 30 minutes, cooled at room temperature and filtered using Whatman filter paper. Then the filtrate was sterilized in an autoclave for 15 minutes for 15 ponds. Later the autoclaved hay filtrate was kept in the lab for 24 hours without any lid. For culturing ciliates, Hay media was diluted in the ratio of 1:1. Cultures were maintained at room temperature $25 \pm 2^{\circ}\text{C}$. In order to induce ciliate multiplication boiled wheat grains were added to culture media. Log phase cultures were used throughout the study.

Acute toxicity studies: Acute toxicity tests were conducted for 3 hours to measure immediate cell response, such as swimming pattern, cell motility and cytopathological changes under pesticidal stress as suggested by Apostol, (1973). The tests were repeated thrice. 0.5 ml of known concentration of deltamethrin solution was added to 4.5 ml of culture medium containing about 50 organisms. Observations were made and counting was done for every 10 min during the first hour and after 20 min interval for next two hours. Total disintegration of the cell was used as the lethal point. Immediate cytopathological responses were recorded at various concentrations to determine the LC_{50} values by plotting a graph (Finney 1964).

Contractile Vacuole Activity: Contractile Vacuole discharges its contents through a permanent pore (Patterson 1981). Immobilization of the *Blepharisma intermedium* was done by using protamine coated slides. As the test organisms appeared not to be harmed by this procedure (Marsot and Couillard, 1973). The organisms were exposed to sub-lethal concentrations 10ppm, 13ppm, 16ppm and 19ppm of deltamethrin for 10 and 20 minutes, single individual organism was picked and the rate of pulsation of contractile vacuole i.e. time required for one complete pulsation was determined. The rate of pulsation for each individual organism was calculated separately. Equal number of observations were done in controls.

Food vacuole activity: About 25 organisms were exposed to sub-lethal concentrations 10ppm, 13ppm, 16ppm and 19ppm of deltamethrin for one hour respectively, to identify the number of food vacuoles formed. The test organisms were divided into two groups, 1) Treated organisms, 2) control organisms. Treated organism from each concentration were picked with the help of micropipette, mixed with India ink and kept for 10 minutes. 10 organisms from each concentration were taken, immobilised and number of food vacuoles formed was recorded. Control cells with same molar concentrations of India

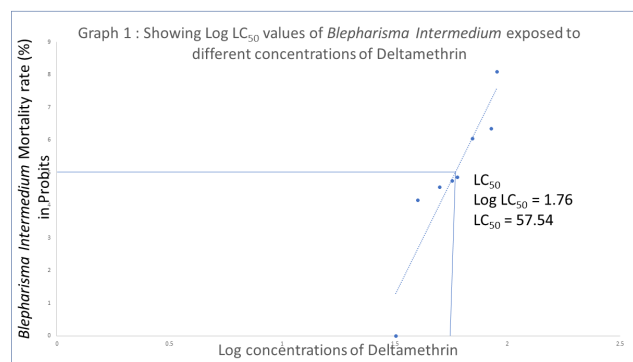
ink was run simultaneously. A few organisms are collected from both the experimental groups on protamine coated slides for immobilization and the counting is done. Preparation of carmine suspension and counting of food vacuoles was done as method suggested by Nilsson (2003).

Macro Nuclear Aberration study

Feulgen fast green technique Feulgen fast green (Rizzo and Nooden ; 1973) staining technique was used to study the macro nuclear changes in *Blepharisma intermedium* exposed to deltamethrin. Macro nuclear studies gives an immediate qualitative picture of the nuclear changes that have been induced by the pesticides (Nageswara Rao Amanchi, 2009). The organisms were treated with sub lethal concentrations of 10ppm, 13ppm, 16ppm and 19ppm for one-hour exposure to deltamethrin. Carnoy's fixative was used for cell fixation. The treated organisms were hydrolysed in 1N HCL at 27°C temperature and washed in distilled water, slides were transferred into Schiff's reagent and incubated for 1 hr. Schiff's reagent was prepared as suggested by De Tomasi (1930). The organisms were immersed in three changes of sodium bisulphate solutions, again rinsed with water, dehydrated in graded alcohol, cleared in xylene and mounted with DPX. The exposed *Blepharisma intermedium* was shown various macronuclear changes like rod shape, vacuolated, unevenly divided, fragmented macronucleus and karyolytic forms were observed.

RESULTS

Acute toxicity studies: *Blepharisma Intermedium* was exposed to different concentrations of deltamethrin for 3 hrs, various changes in the body size, shape, ultrastructural deformities were observed. The lethal point was found at the concentration of 90ppm for *Blepharisma Intermedium* (graph: 1) and the calculated LC_{50} value against mortality curve was found to be 57.54ppm. The worked out sublethal concentrations for *Blepharisma Intermedium* were 10ppm, 13ppm, 16ppm and 19ppm respectively.



Graph 1 showing Acute toxicity effect of deltamethrin on *Blepharisma Intermedium*.

Contractile Vacuole Activity: Contractile vacuole activity was performed for each individual organism after exposing 10ppm, 13ppm, 16ppm and 19ppm concentrations for 10 & 20 minutes of exposure time. Observations were made for individual cells by comparing with control. There was maximum reduction in number of pulsations of *Blepharisma Intermedium* (table 1) was found to be 1.00 ± 0.63 , 0.80 ± 0.40 and 1.40 ± 0.48 , 0.80 ± 0.74 at 19ppm and 16ppm

concentrations of Deltamethrin for 10 & 20 minutes. compared to control organism 3.80±0.40. The minimum reduction 2.20±0.40 and 1.60±0.48 observed at the concentration of 10ppm in treated cell of the *Blepharisma Intermedium*. (One-way ANOVA was done $F_{4,20} = 22.296$; $P < 0.05$).

Table 1 Contractile activity in *Blepharisma Intermedium* exposed to Deltamethrin

Concentration in ppm	Exposure time	Pulsations per minute
10	10	2.20±0.40
	20	1.60±0.48
13	10	2.00±0.63
	20	1.20±0.40
16	10	1.40±0.48
	20	0.80±0.74
19	10	1.00±0.63
	20	0.80±0.40
Control		3.80±0.40

One way ANOVA was done $F_{4,20} = 22.296\%$; $P < 0.05$

Food vacuole activity: *Blepharisma intermedium* shows reduction in number of food vacuoles (fig: 1) against different concentrations of Deltamethrin. The gradual decrease in the phagocytic activity was recorded by increasing the concentration. Minimum inhibition of food vacuole was for one exposure 34% at concentration of 10ppm. Maximum inhibition was found 76% at concentration. (one-way ANOVA showing $F_{4,45} = 93.603$; $P < 0.05$).

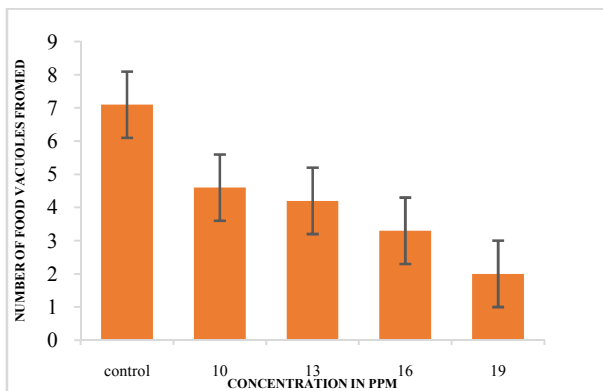


Fig 1 Food vacuole activity in *Blepharisma intermedium* exposed to different concentrations of Deltamethrin for 1hr. Cytochemical changes of nucleus

The treated groups of *Blepharisma Intermedium* (Table 2) has shown various macronuclear changes including vacuolated, fragmented, unevenly divided, karyolysis. Percent abnormal forms reported were 23.2±0.63, 34.1±0.74, 51.6±0.7 and 67.2±0.79 at each concentration of 10ppm, 13ppm, 16ppm and 19ppm. Presence of maximum number of vacuolated forms were recorded were 20.7±0.67 at 19ppm and the minimum values recorded were 6.6±0.7 at concentration 10ppm.

Table 2 Deltamethrin induced nuclear changes (%) in *Blepharisma Intermedium* exposed for one hour.

Conc/ppm	Percent abnormal forms	Vacuolated	Fragmented	Unevenly divided	karyolysis	Other deformities
10	23.2±0.63	6.6±0.7	8.3±0.82	3.5±0.71	10.4±0.52	4.4±0.7
13	34.1±0.74	7.3±0.67	9.4±0.52	3.8±0.63	13.4±0.7	7.3±0.67
16	51.6±0.7	12.2±0.63	13.8±0.79	6.3±0.67	12.4±0.52	10.8±0.79
19	67.2±0.79	20.7±0.67	18.6±0.52	11.1±0.88	18.4±0.52	11±0.67
Control	2.7±0.67	---	2±0.47	---	---	1.4±0.521

P values are significant $P < 0.05$

DISCUSSION

The use of insecticides has increased due to pest control management in agriculture and household applications. Pyrethroids are neurotoxic not only to insects but also to the mammals and zooplankton including fresh water ciliates. Pyrethroids interact with sodium channels of nervous system and inactivates sodium current following by membrane depolarisation (Narahashi, 1992). Low concentrations stimulate the nerve membrane and at higher concentrations block the nerve entirely (Rose *et al.*, 2001).

Acute toxicity studies were carried out to observe the immediate cytopathological responses. Changes in the body size, shape and ultrastructural deformities were observed in ciliates, when exposed to different concentrations of deltamethrin. The data obtained from the acute toxicity studies was important in establishing the relative toxicity of pesticides, as well as in providing information to carry out further experiments. Studies reveal that the action of deltamethrin on both calcium uptake and depolarization-evoked neurotransmitter release in rat brain (Symington *et al.* 2007b; Symington *et al.* 2008). Similar results were also reported by various authors (Komala 1982, Schultz *et al.* 1981, Masood, Amanchi 2008) in different experimental organisms.

Blepharisma Intermedium showed maximum reduction in the contractile vacuole activity when the cells were exposed to higher concentrations of 18ppm and 19ppm of deltamethrin. The recorded results were found to be 1.00 ± 0.63 , 0.80 ± 0.74 and 2.20 ± 0.40 , 1.60 ± 0.80 for 10 & 20 minutes, due to the effect of deltamethrin. Minimum reduction was observed at lower concentrations of deltamethrin. The effect of toxicant was studied by (Rossbach 1872; Hartog 1888). If the osmotic pressure of the medium is increased, expulsion of the contractile vacuole decreases. If the added toxicant is diluted the expulsion will increase. (Stoner & Dunham, 1970; Hampton & Schwartz, 1976). The ion physiological studies influence the functions of contractile vacuole (Dunham & Child (1961). Amoeba and Tetrahymena have shown reduced contractile vacuole activity when the cells are subject to a hyperosmotic shock (Prusch (1977) and Dunham & Kropp (1973).

Damage to cilia and ultra-structure of plasma membrane causes reduction in food vacuole formation. The cilia present in cytostome may get damaged under the insecticidal influx into cytoplasm from outside. Under stress conditions ciliates exhibit starvation to avoid unfavorable conditions (Khan and Masood; 1983). More number of nuclear abnormal forms were observed when organisms were exposed to various sublethal concentrations of deltamethrin. Reduction in size, shape, rod shape, vacuolated, blebbing and blacking of cytoplasm were observed at higher concentrations. *Blepharisma Intermedium* exposed to the concentration of 19ppm the percent abnormal forms were found to be (67.2 ± 0.79) . Higher concentrations the ciliary movement was arrested and lysis of the organism was observed.

CONCLUSION

Commercial grade deltamethrin was toxic even at very low concentrations like 10ppm, 13ppm, 16ppm and 19ppm. Changes in osmotic media influenced both contractile vacuole

activity and food vacuole activity in *Blepharisma Intermedium*. A beaded nucleus of *Blepharisma Intermedium* is highly useful in studying genotoxicity effects of deltamethrin. Hence it is concluded that *Blepharisma Intermedium* is a suitable model for basic bioassay studies as well as mutagenic and carcinogenic studies.

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