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**RESEARCH ARTICLE**

**EFFECT OF TWOTRICHODERMA HARZIANUM SPECIES IN FUSARIUMSOLANI AND CYLINDROSPORIUMSP, PATHOGENS OF LENS CULINARIS**

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**ABSTRACT**

The aim of this study was to evaluate in vitro inhibitory activity of Trichoderma harzianum 127a and 127b Trichoderma harzianum. For this, two kinds of samples were made: of soil samples at different depth (5, 10 and 15 cm), and samples of the different plant organs (roots, stems and leaves). Direct and indirect confrontation techniques have served tested for different metabolites. While the technique has served, inserts evaluated the effect of the opposing competitor strains. The results of the various confrontations showed that the inhibition rate was around 90% for the two antagonistic strains T-127a and 127b-T. Furthermore, comparison tests showed a significant reduction in germination. The two strains of Trichoderma harzianum tested in this study showed a high efficiency vis-à-vis Cylindrosporium sp and Fusarium solani pathogenic lenses in the Constantine region.

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**INTRODUCTION**

The presence of fungal diseases on Lens culinaris plant and its economical consequences require the use of many pesticides. Chemical pesticides that are widely used to control plant pathogens do not degrade completely and leave toxic residues in food chain (Chet, 1987; Lynch, 1990). However, biological control agents to fight plant diseases become more and more widespread. Several Trichoderma species are recognized from 1930 as biocontrol (Biljana Gveroska and Jugoslav Ziberoski, 2012) and today there are modern technologies for including them in the biological control of various diseases.

Trichoderma was described in 1794, including anamorphic fungi isolated primarily from soil and decomposing organic matter (Persoon, 1794), and approximately 130 species have been classified (Degenkolb, 2008). Trichoderma free-living fungi that are highly interactive in root, soil and foliar environments. The most commonly used strains of Trichoderma bio control agents are Trichoderma viridescens, Trichoderma viride and Trichoderma harzianum (Grondona I. and et al., 1997).

The strains of Trichoderma used as biocontrol agents show different mechanisms of action in their antagonistic interactions with fungal pathogens. These include antibiosis, through the production of a variety of compounds with antibiotic activity (Ghisalberti and Sivasithamparam, 1991), mycoparasitism or hyperparasitism (Ayers and Adams, 1981), competition for nutrients (Chet, 1987), cell wall-lytic enzyme activity (Lorito et al., 1996; Lorito 1998) and induction of systemic resistance to pathogens in planta (Harman and et al., 2004).

Results from different studies showed that several strains of Trichoderma exerted significant reducing effects on plant diseases caused by pathogens such as Rizoctonia, Pythium, Fusarium, Gaeumannomyces under greenhouse and field conditions (Sivan and Chet 1993; Chet and Baker 1981). According to Rosado et al. (2007), the main factor for ecological success of this genus is a combination of very active mycoparasitism mechanisms and an effective defensive strategy, induced in the plants. Our work is oriented to study the effect of two Trichoderma species in Fusarium solani and Cylindrosporium pathogens of Lens culinaris.

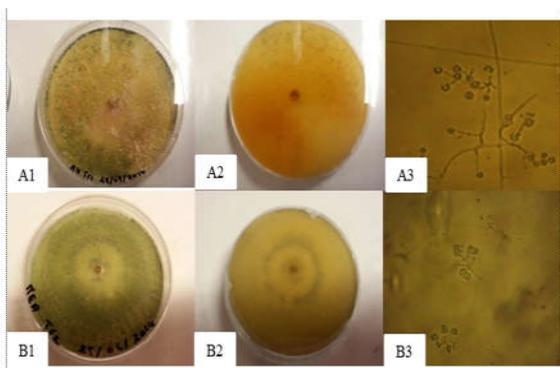
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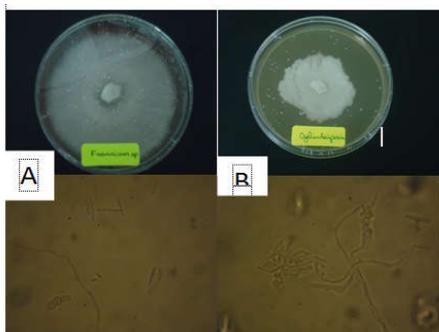
## MATERIAL AND METHODS

### Screening of *Trichodermaharzianum* from agriculture soil

Soil samples taken at 5, 10 and 15 cm in depth were collected from four locations in the vicinity of the two areas (AIN SEMARA and BARAOUIA), in sterile paper bags. The soils collected from each area were mixed thoroughly and *Trichodermaharzianum* spores were isolated using dilution plate and soilplate techniques on PDA and malt extract agar (MEA) with antibiotics.



**Fig 1** The two strains of *Trichoderma*: (A): *Trichoderma harzianum* 127a, (B): *Trichoderma harzianum* 127b- Les deux souches antagonistes de *Trichoderma* : (A): *Trichoderma harzianum* 127a, (B): *Trichoderma harzianum* 127b.



**Fig 2** Macroscopic and microscopic characteristics of *Fusarium solani* (A) and *Cylindrosporium* sp (B)-Caractères macroscopiques et microscopiques de *Fusarium solani* (A) ET *Cylindrosporium* sp (B)

### Isolation of pathogens spores from plants

*Fusariumsolani* and *Cylindrosporiumsp* pathogens of *Lens culinaris* were isolated from different hosts showing typical disease symptoms. Isolation of pathogens was done by cutting 0.5 cm pieces of diseased samples and surface sterilizing them in with Sodium Hypochlorite (2%) for 3 min, then rinsed several times with sterile distilled water and ethanol (70%) and dried between sterile filter paper. The pieces treatment was transferred to sterilized PDA in petriplates. The Petriplates were incubated at 27 ° C for 4 days (Belabid and et al., 2000). Finally, myceliums obtained are purified in sterilized malt extract agar (MEA).

### Identification of spores

#### Microscopy

Strains isolated and purified in malt extract agar were

examined under optic Zeiss Axioskop 2 MOT microscope where morphology was noted besides hyphal structure, spore size, shapes and spores bearing structures. They were compared with the standard works of Samson et al Haesks (1988); Hawkswarth et al. (1994); HoogandGurro (1995) and Gams et al. (1998).

### Identification by sequencing segment of the *Tef 1-α* gene

The two strains of *Trichoderma* isolated were identified in ULC (Catholic University of Louvain-La-Neuve- Belgium). The DNA of the segment of the *Tef 1-α* gene was sequenced and compared with reference sequences deposited in public database, cross checked with the morphology.

### Antagonist tests

#### Diffusible metabolites tests (DM Tests)

The effect of non-volatile metabolites from *Trichoderma harzianum* species against *Fusarium solani* and *Cylindrosporiumsp* was tested by the method described by Dennis and Webster (1971), and Lundberg and Unestan (1980). This technique consists of placing, in the same Petri dish containing PDA medium, two agar pellets (6 mm diameter), one bearing the *T. harzianum* and the other gold *C. sp* or *F. solani*. The two wafers are positioned along a diametral axis to 3cm and equidistant from the center of the box; subcultures are performed simultaneously. Incubation was carried out at 30°C for six days. Colony size in each treatment was recorded and percent inhibition was calculated by using the formula as proposed by Vincent (1947).

$$I = (C - T / C) \times 100.$$

Where:

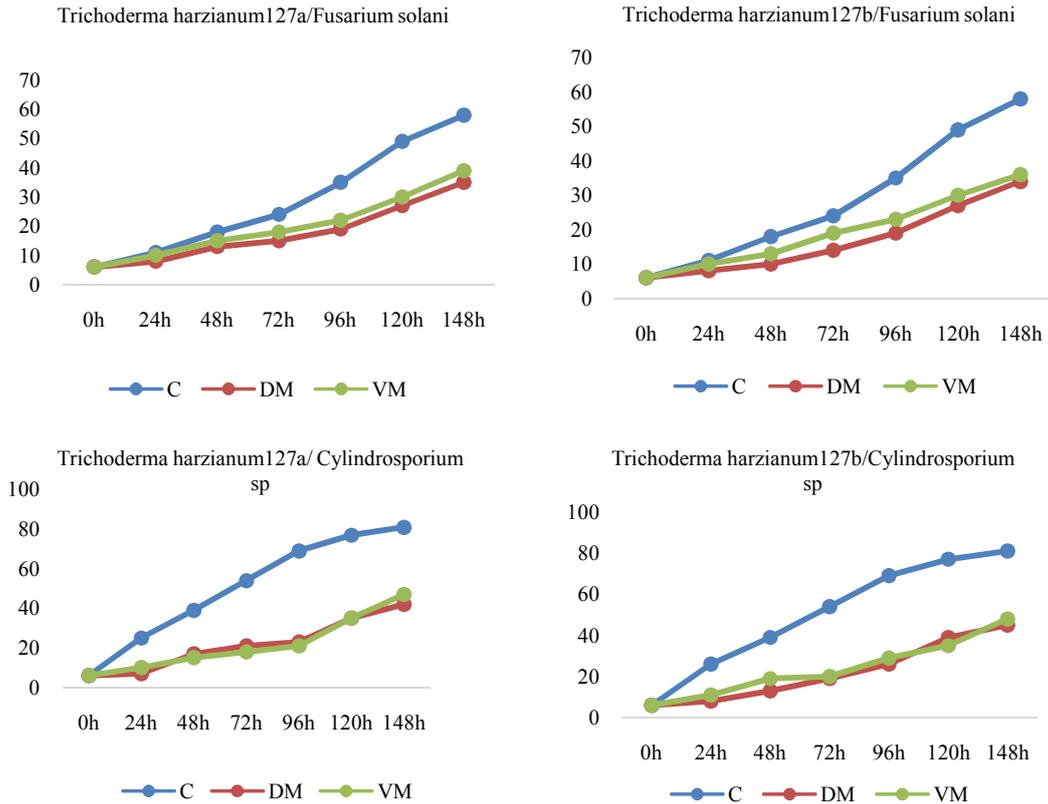
- ❖ I = Inhibition of mycelial growth (%).
- ❖ C = Growth of pathogen in control (mm).
- ❖ T = Growth of pathogen in treatment (mm).

#### Volatile metabolites tests (VM Tests)

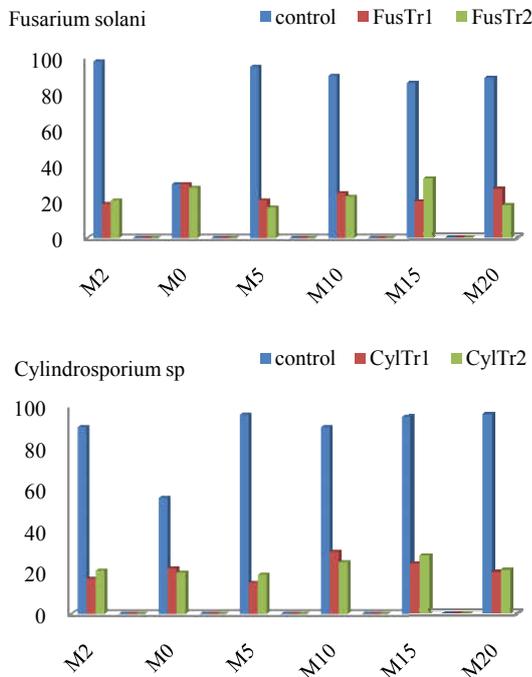
In this technique, the two pellets (6mm diameter) (as I, Diffusible metabolites tests), are placing in two different Petri dishes containing PDA medium. The two Petri dishes are sealed with parafilm in a spetic condition and incubate for six days at 30°C. The percent of inhibition was calculated as in the last test.

### 5- Study of competition activity of *Trichoderma* In vitro

Competition for nutrition was studied in vitro by using a non-destructive method developed by Janisiewicz and et al. (2000). The device consists of cell culture 24-well plates (TC-tests plats) and of cylindrical inserts (BD Falcon Cell Culture Inserts, 0.4µm). In this method, the antagonist effect of *Trichodermaharzianum* on *Cylindrosporiumsp* and *Fusarium solani* was tested at different glucose concentration of MEA medium (0, 5, 10, 15, 20 g / L) and M2 medium. The effect of strains of *T. harzianum* on *C. sp* and *F. solani* in the different environments was evaluated by comparing the percentage of germination of conidia in each case with the corresponding control.



**Fig 3** Inhibition of mycelia growth of *Fusarium solani* and *Cylindrosporium* sp by diffusible metabolites (DM) and volatile metabolites (VM) produced by T-127a and T-127b- Inhibition de la croissance de *Fusarium solani* et de *Cylindrosporium* sp par les métabolites diffusibles (DM) et les métabolites volatiles (VM) produits par T-127a et T-127b.



**Fig 4** Percentage of conidial germination of *Fusarium solani* and *Cylindrosporium* sp in the absent and present of antagonists T-127a and T-127b- Pourcentage de la germination des conidies de *Fusarium solani* et de *Cylindrosporium* sp en présence et en absence de T-127a et T-127b.

## DISCUSSION

The idea of a sustainable agricultural practice and environmental protection enhances the importance of biocontrol. One of the key elements of such sustainable agriculture is the application of biocontrol agents (Cook and *et al.*, 1989; Deacon and *et al.*, 1992). The genus *Trichoderma* is most important in achieving that and, at the same time, sustaining a favourable environment, instead of using chemicals.

The results obtained during the various confrontations (antagonism tests: VM, DM) conducted *in vitro*, showed a highly significant inhibition (60 to 92%) of pathogens growth during the study. Indeed, species of *Trichoderma* develop, exactly on other fungi's hyphae, coils around them and degrade the cell's walls. This action of parasitism restricts the development and activity of pathogenic fungi. Additionally, or together with mycoparasitism, some *Trichoderma* species release antibiotics (Harman, 1996). When the *Trichoderma harzianum* 127a and T-127b, are placed in Petri dishes, they have a space advantage to develop rapidly and to stop pathogen development, and to develop its mechanisms of antagonistic action. In a short time, it significantly reduces growth of *Fusarium solani* and *Cylindrosporium* sp. The pathogen develops until the moment of contact with the antagonist; simultaneously it sporulates and changes the colony's color, acting as a super-parasite (Mirkova, 1983). Küçük and Kivanç (2003) showed that in general, the diffusible

metabolites have a bigger reducing effect than the volatile ones.

In vitro-made competition tests showed that in the absence of antagonists strains (control), the germination of conidia of *Fusariumsolani* and *Cylindrosporiumsp* was better in the presence or absence of glucose in the medium. This confirms that *F.solani* and *C. sp* are dependent nutrient environment and need enough nutrients to germinate and become infectious. Indeed, *Trichodermaharzianum*127a and *T.harzianum* 127b has grown faster than *Fusariumsolani* and *Cylindrosporiumsp*, in MEA culture. This rapid growth gives *Trichoderma* species an important advantage in the competition for space and nutrients with plant pathogenic fungi (Barbosa, Rehm *et al.*, 2001).

The efficacy of the genus *Trichoderma* strains, and especially those belonging to the genus *Trichodermaharzianum*, was also proved by the different results of studies of Basin H. *et al.* (1999), Faheem Amin *et al.* (2010), Inbar *et al.* (1994), Sivan and Chet (1993) and Ubalua *et al.* (2007), which reported that *Trichoderma* has significant inhibitory effects on pathogenic strains which *Fusariumsolani* and *Cylindrosporium sp*.

## CONCLUSION

This study confirms In vitro biological activity of *Trichodermaharzianum* 127a and *T- harzianum* 127b towards *Fusarium solani* and *Cylindrosporium sp*. It has also shown an important reduction effect on the development of these pathogens of *Lens culinaris*. The diffusible metabolites have shown more reducing effects than the volatile ones.

## Knowledge

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## References

1. Belabid L., Fortas Z., Dalli D., Khiare D et Amjad D.(2000). Flétrissement *et* pourriture racinaire de la lentille dans le Nord-Ouest Algerien. Cahiers Agricultures, Volume 9, Numéro 5, 518-8. Notes de recherche.
2. Biljana Gveroska and JugoslavZiberoski. (2012). *Trichoderma harzianum* as a biocontrol agent against *Alternariaalternata*on tobacco. Applied Technologies & Innovations Volume 7 .Issue 2 ; pp.67-76.
3. Barbosa, M., Rehm, K., Menezes, M., Mariano, R.L.(2001). "Antagonism of *Trichoderma* species on *Cladosporiumherbarum* and their enzymatic characterization," Brazilian *Journal of Microbiology*, Vol.32, pp.98-104
4. Basin H., S.B. Ozturk and O. Yegen.(1999). "Efficacy of a biological fungicide (Planter Box *Trichoderma harzianum* Rifai (T=22) against seedling root rot pathogens (*Rhizoctoniasolani*, *Fusariumsp*. Of cotton," GAP-Environmental Symposium, Sanleurfa, Turkey. pp. 137-144.
5. Chet I.(1987). *Trichoderma* applications, mode of action and potential as a biocontrol agent of soilborne plant pathogenic fungi. In Chet I, (Ed.). Innovative approaches to plant disease control. John Wiley & Sons, New York. N.Y. pp. 137-160.
6. Chet I and Baker R. (1981). Isolation and biocontrol potential of *T. hamatum* from soil naturally suppressive to *R. solani*. *Phytopathol*71: 286-290.
7. Chet, in: I. Chet (Ed.). (1987).Innovative approaches to plant disease control, Wiley, New York, pp 137-160.
8. Cook, R.J.; Baker, K.F. (1989).The nature a practice of biological control of plant pathogens. APS Press, St. Paul, 539p.
9. Deacon, J.W.; Berry, L.A. (1992). Modes of actions of mycoparasites in relation to biocontrol of soilborne plant pathogens. In: Tjamos, E.C.; Papavizas, G.C.; Cook, R.J. (eds). *Biological Control of Plant Diseases*. Plenum Press, New York.p.157-167.
10. Degenkolb, T., H. V. Dohren, N. F. Nielsen, G. J. Samuels, and H. Bruckner. (2008). Recent advances and future prospects in peptaibiotics, hydrophobin, and mycotoxin research, and their importance for chemotaxonomy of *Trichoderma* and *Hypocrea*. *Chem. Biodivers.* 5: 671-680.
11. Dennis, C.; Webster, J..(1980). Antagonistic properties of species-groups of *Trichoderma*. I. Production of non-volatile antibiotics. *Trans. Br.Mycol. Soc.*, 84:25-39, 1971a.
12. Faheem Amin, V. K. Razdan, F. A. Mohiddin, K. A. Bhat and P. A. Sheikh. (2010). Effect of volatile metabolites of *Trichoerma* species against seven fungal plant pathogens in vitro. *Journal of Phytology* 2010, 2(10): 34–37.
13. Gams W. Haekstra E.S. and Aptroot A..(1998). CBS. Course of mycology. Central lureauvoor. Schimmel culture Baarns. The Netherlands.
14. Grondona I., M.R. Hermosa, M. Tejada, M.D. Gomis, P.F. Mateos, P.D. Bridge, E. Monte and I. Garcia-Acha. (1997). Physiological and biochemical characterization of *Trichoderma harzianum*, a biological control agents against soil borne fungal plant pathogens. *Appl Environ Microbiol*, vol. 63, pp. 3189-3198.
15. Harman, G.E..(1996). "Trichoderma for biocontrol of plant pathogens: From basic research to comercialized products," Conference on Biological Control, Cornell community, April 11-13
16. Hawksworth D.L., Kirk P.M., Sutton B.C. and Pegler D.N..(1994). Ainsworth and Bysby's dictionary of the fungi, 8 thed . International Mycological Institute, Egham. Unitted .Kingdom.
17. Hoog G.S., GUARRO J..(1995). Atlas of clinical fungi, central bureauvoor Scimmel cultures, Baar. Pays-Ba.
18. Inbar J, Abramsky M and Cohen D..(1994). Plant growth enhancement and disease control by *T.harzianum* in vegetable seedlings grown under commercial conditions. *European J PlaPathol*100:337-346.
19. Janisiewicz W.J., Tworowski T.J., and Sharer C..(2000). Characterizing the mechanism of biological control of postharvest diseases on fruits with a simple method to study competition for nutrients. *Phytopathology*, 90 (11), 1196-1200.

20. Lundberg, A.; Unestan, T.. (1971). Antagonism against *Fomesannosus*. Comparison between different test methods "in vitro" and "in vivo". *Mycopathologia*, 70:107-115.
21. Lynch JM. (1990). Fungi as antagonists In: *New directions in biological control: Alternatives for suppressing agricultural pests and diseases*, Liss, New York, pp. 243-253.
22. Mirkova, E. (1983). "Priloženiena *Trichoderma harzianum* Rifaizaborba s fuzariinotouvrhvane (*Fusarium oxysporum*sch.f. *dianthi* (Prill. Et Del.) Snyd. Et Hans.) prioranzeriniakaramfil," *Selskostopanskaakademia, Gradinarskailozarskanauka, god. XX, No.1, pp.65-69*
23. Rosado, I., Rey, M., Codon, A., Gonavites, J., Moreno-Mateos, M.A., Benitez, T..(2007). "QID74Cell wall protein of *Trichoderma harzianum* is involved in cell protection and adherence to hydrophobic surfaces," *Fungal Genetics and Biology*, Vol.44(10), pp.950-64
24. Samson R.A. and Hoekstra E.S. (1988). *Introduction to food-borne fungi*, 3rd edn. Central bureau Voor. Schimmel cultures. Baane. The Netherlands.
25. Sivan A and Chet I. (1993). Integrated control of media on growth and interactions between a range of soilborne glasshouse pathogens and antagonistic fungi. *Phytopathology* 10: 127-142.
26. Ubalua, A. O. and Oti, E. (2007). Antagonistic properties of *Trichoderma viride* on post harvest cassava root rot pathogens. *African Journal of Biotechnology* Vol. 6 (21), pp. 2447-2450,
27. Vincent, J.M. and Budge, S.P. (1990). Screening for sclerotialmyco parasites of *Sclerotinia sclerotiorum*. *Mycological Research* 94: 607-612.
28. Ghisalberti E.L. and Sivasithamparam K. (1991). Antifungal antibiotics produced by *Trichoderma* spp. *Soil. Biol. Biochem.* 23 : 1011-1020.
29. Ayers W.A. and Adams P.B.(1981). Biological control of *Sclerotinia* lettuce drop in the field by *Sporidismium sclerotivocum*; *Phytopathology*. P: 485-9.
30. Lorito M., Hayes C.K., Di Pietro A., Woo S.L., Harman G.E.(1994). Purification, characterization and synergistic activity of a glucan 1,3-b-glucosidase and an N-acetyl-b-glucosaminidase from *Trichoderma harzianum*. *Phytopathology* 84: 398-405.
31. Lorito M., Hayes C.K., Di Pietro A., Woo S.L., Harman G.E. (1994). Purification, characterization and synergistic activity of a glucan 1,3-β-glucosidase and an N-acetyl-β-glucosaminidase from *Trichoderma harzianum*. *Phytopathology*. 84: 398-405.
32. Lorito M., Woo S.L., Garcia Fernandez I., Colucci G., Harman G.E., Pintor-Toro J.A., Filippone E., Muccifora S., Lawrence C.B., Zoina A., Tuzun S., Scala F., (1998). Genes from mycoparasitic fungi as a novel source for improving plant resistance to fungal pathogens. *Proceedings of the National Academy of Sciences USA* 95: 7860-7865.

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