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Research Article

STUDIES ON BACTERIAL, FUNGAL DIVERSITY OF KUMBAKONAM TALUK THANJAVUR DISTRICT, TAMIL NADU, INDIA

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ABSTRACT

Soil are highly complex system, with many component playing diverse functions mainly due to the activity of soil organism. Soil flora plays a pivotal role in evaluation of soil condition and in stimulating plant growth. Microorganism are beneficial in increasing the soil fertility and plant growth as they are involved in several biochemical transformation and mineralization the activities in soils. The present study was aimed to, deals with the diversity at sites in Kumbakonam Taluk, Thanjavur District, TamilNadu. The study period was covering all the four seasons viz, Monsoon (October-December), Postmonsoon (January- March) Summer (April – June) Premonsoon (July – September) and distribution of bacterial and fungal population in around soil. The physico-chemical parameters of such soil were identified the includes pH and moisture content of the soil. macronutrient (Nitrogen, Phosphorus, Magnesium, Calcium) and micronutrient (Iron, Copper, Zinc, Manganese) were analyzed. Totally 50 species of soil bacteria and fungi were observed from the soil samples, they were collected from Kumbakonam, Krishnapuram and Cholapuram. About 30 different species belonging to Ascomycetes and Phycomycetes and most of them were isolated by using PDA medium and Nutrient Agar Medium identified with standard manuals. The dominant bacterial species are *Pseudomonas sp*, *Clostridium sp*, *Bacillus sp*, *Staphylococcus sp* and *Escherichia sp* and fungal species *Aspergillus sp*, *Rhizopus sp*, *Pencillium sp*, and *Fusarium sp* were recorded. Isolation of microorganisms were correlated to percentage frequency and heavy metal content were recorded. The physic –chemical parameters are rich in Kumbakonam as Silty Clay to Silty Clay Loam soil type.

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INTRODUCTION

Soil is a complex ecosystem, delimited by physiochemical parameters that hold enormous number of living organism. Nevertheless, microbes are the least understand mechanism of soil by both agronomists and soil practitioners. On the farm several soil organisms offer benefits to crop growing in an ecosystem, but are not well understood. The microbes decompose the plant and animal residues entering the soil and convert them into soil organic matter, which influences on soil physical, chemical and biological properties and on creating a complimentary medium for biological reaction and life support in the soil environment. Nonetheless, enhanced site-specific diversity typically results in higher levels of below ground microbial diversity and production (Olson *et al.*, 2000). Biodiversity refers to the variability of life on Earth, all the living species of animal, plant and microorganism. According to Hawksworth (2001), fungi are a major component of biodiversity, essential for the survival of other organisms and

are crucial in global ecological processes. Biodiversity is not evenly distributed rather it varies greatly across the globe as well as within region. Among other factor, the diversity of all living thing (biota) depends on temperature, precipitation, altitude, soils, geography and the presence of other species.

Soil biodiversity reflects the mix of living organism in the soil. These organism interact with one another and with plants and animals forming a web of biological activity. Soil is by far the most biologically diverse part of earth. The soil food web includes beetles, springtails, mites, worms, spiders, ants, nematodes, fungi, bacteria, and other organisms. These organisms improve the entry and storage of water, resistance to erosion, plant nutrition, and break down of organic matter. A wide variety of organisms provides check and balances to the soil food web through population control, mobility, and survival from to season. Climate change affect Biodiversity (2014). Hence, the present study was isolate and identify bacteria and fungi in Thanjavur Dt, Tamilnadu during four different seasons.

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MATERIALS AND METHODS

Soil Sample Collection

The soil samples were collected from Kumbakonam taluk, Thanjavur District, Tamil Nadu, India during Monsoon (October-December), Post monsoon (January- March) Summer (April – June) Premonsoon (July – September (2014-2015) season. In the present case, each sample was collected agricultural field, and from taluk namely Kumbakonam (S1), Krishnapuram (S2) and Cholapuram (S3). Collect the soil sample before remove the surface litter at the sampling spot. Collect at least 4 to 6 samples from each sampling unit and place. If make a 'V' shaped cut to a depth of 15 cm in the sampling spot using spade.

Mix the sample thoroughly and remove foreign material like roots, stones and gravels. Silty clay to. Silty clay loam the method used for taking soil sample was a slight modification as that used by (Goddard 1913). Collect the in sample clean sterile polythene bags. All four random samples of each zone were put together to make a single sample from each place. A total of three samples was prepared to investigate the diversity of the bacteria and fungi.

Isolation of Bacteria and Fungi

Isolation was done by serial dilution and dilution plating (Gram's Staining, Motility) using and standard manuals (Bergey's manual of determinative bacteriology) and Identification in terms of biochemical test such as Indole, Methyl – Red and Voges- Proskauert, Citrate Utilization, Catalase, and Oxidase Test. The bacterial and fungal colonies were counted as cfu/ ml.

Physico chemical properties (Griffin, 1970)

The pH, moisture, organic carbon Macronutrients (Nitrogen, phosphorus, Manganese, Calcium) and micronutrients (Iron, Zinc, Manganese) were analysed and compared with seasonal variations of per standard method. (Table 4)

Estimation of Trace Elements (Piper, 1944)

Copper, Iron, Manganese and Zinc heavy were analyzed by using Atomic Adsorption Spectrophotometer. (AAS Elico 196).

Procedure

Filter the samples and acidified with concentrated HNO₃ to a pH less than immediately after collection of soil.

Take 1.5 liter of filtrated sample was taken in a beaker with 5ml concentrated HNO₃. Evaporated the sample was dried on a hot plate to prevented the boiling process. Add another 5 ml of the concentrated HNO₃ in the sample. Then it allow to cooled and heated by adding some additional HNO₃ light colored residue were formed. Dissolve the residues by added 0.5N HCL and filtered the contents. Makeup the filtered content upto 50 ml with 0.5N HCL. After pretreatment the minerals can be analyzed by Atomic Adsorption Spectrophotometer.

Statistical Analysis (Salil bose 1982)

The results obtained in the present investigation were submerged to statistical analysis like mean (X) and Standard Deviation (SD).

RESULTS AND DISCUSSIONS

The present study was aimed to investigated for the bacterial and fungal diversity from Kumbakonam taluk of Thanjavur district, sites such as, Kumbakonam, (S₁) Krishnapuram (S₂) and Cholapuram (S₃). Tamilnadu in monsoon, postmonsoon, summer, and premonsoon seasons.

Physico- chemical parameters

The pH, moisture, organic carbon Macronutrients (Nitrogen, phosphorus, Manganese, Calcium) and micronutrients (Iron, Zinc, Manganese) were analysed and compared with seasonal variation of Kumbakonam taluk of Thanjavur District. (Table 4)

The bacterial and fungal organisms are isolated and identified as morphological and cultural characteristics (Table 9, 10). Nearly 32 species (bacteria 12 and fungi 20) were recorded in monsoon, 48 species (bacteria 14 and fungi 21) were recorded in post monsoon, 31 species (bacteria 16 and fungi 15) were recorded in summer and 47 species (bacteria 18 and fungi 29) were recorded in Premonsoon season. (Table 1). Bacterial and fungal colonies were counted and recorded as cfu/ml (Table 5,6).

Table – 1 Isolation of bacteria and fungi from soil during four season

S.No	Species	Seasons			
		Monsoon	Postmonsoon	Summer	Premonsoon
1	Bacteria	12	14	16	18
2	Fungi	20	21	15	29

In monsoon season, in the area of Kumbakonam (S₁) site, fungal organisms are *Aspergillus niger*, *A. flavus*, *A. nidulans*, *A. sulphureus*, *Penicillium sp.*, *p. chrysogenum*, *P. bovis*, *R. oryzae*, *C. herbarum*, *Mucor sp.*, *Alternaria sp.*, *Fusarium solani*, *Phythium sp.*, *Verticillium sp.*, *R. nigricans* and bacterial species such as *Bacillus sp.*, *Enterobacter sp.*, *P. vulgaris*, *Streptococcus sp.*, *E. aerogenes*, *S. lactis*, *Azospirillum sp.* In Krishnapuram (S₂) fungal organisms are *Rhizopus oryzae*, *Candida albicans*, *A. luchuensis*, *Aspergillus flavus*, *C. herbarum*, *P. bovis*, *Fusarium solani*, *F. semitectum* and bacterial species such as *Bacillus cerus*, *B. subtilis*, *Enterobacter sp.*, *E. aerogenes*, *Pseudomonas sp.*, *P. aeruginosa*. Next sites Cholapuram (S₃) fungal organisms are *R. oryzae*, *Aspergillus flavus*, *A. itaconicus*, *A. nidulans*, *A. sulphureus*, *Candida albicans*, *Penicillium chrysogenum*, *Mucor*, *Alternaria sp.* and bacterial species *Micrococcus sp.*, *P. vulgaris*, *Azotobacter*, *Pseudomonas sp.*, *P. aeruginosa* and *Staphylococcus sp.* are dominants.

A review on post monsoon season S₁ site species are fungal *R. oryzae*, *Aspergillus flavus*, *A. itaconicus*, *A. nidulans*, *A. sulphureus*, *Candida albicans*, *Penicillium chrysogenum*, *Mucor*, *Alternaria sp.* and bacterial species *Micrococcus sp.*, *P. vulgaris*, *Azotobacter*, *Pseudomonas sp.*, *P. aeruginosa*, *staphylococcus sp.* In S₂ site fungal organisms are *A. flavus*, *A. fumigatus*, *A. itaconicus*, *Rhizopus sp.*, *R. oryzae*, *C. herbarum*, *Fusarium sp.*, *F. solani*, *Mucor sp.*, *R. nigricans* and bacterial species *Enterobacter sp.*, *E. aerogenes*, *P. vulgaris*, *Pseudomonas sp.*, *P. aeruginosa*, *Lactobacilli sp.* In S₃ site fungal organisms *A. nidulans*, *A. fumigatus*, *A. luchuensis*, *Penicillium sp.*, *P. chrysogenum*, *Mucor sp.*, *Candida albicans*, *R. oryzae*, *Fusarium sp.*, *F. oxysporum* and bacterial species *E. coli*,

Micrococcus sp, *Staphylococcus sp* *B. subtilis* *P.aeruginosa*, *Azospirillum* are dominants

Next Summer season S1 site fungal organisms *A.niger*, *A.fumigatus*, *A.itaconicus*, *Rhizopus sp* *Candida albicans* *R.stolonifer*, *Verticillium sp* *R.nigricans*, *Fusarium oxysporium*, *F. semitectum*, *T. viridae*, and bacterial species are *E.coli*, *S.lactis*, *E. aerogenes*, *Micrococcus sp*, *Aerococcus sp* *Azotobacter P.aeruginosa*. In S2 site species of fungal are *A.niger*, *A.flavus*, *A. sulphurous*, *A. luchuensis*, *Rhizopus sp* *R.oryzae* *R.nigricans* *Fusarium sp*, *F.oxysporum* *F.solani*, *T.viridae* and *Enterobacter sp*, *Pseudomonas sp*, *P.vulgaris*, *Lactobacillus sp* *B. subtilis*. *P.aeruginosa* and S3 site *A.niger*, *A.flavus*, *C. herbarum*, *A.nidulans* *A.itaconicus*, *Candida albicans*, *Pencillium bovis*, *Verticillium sp*, *P.chrysogenum*, *Mucor sp*, *Fusarium solani*, *F.oxysporum*, bacterial species are *E. aerogenes*, *E.coli*, *Micrococcus sp*, *Staphylococcus sp* *S.lactis* are dominants.

In Premonsoon season S1 site fungal species are *Aspergillus A. flavus* *A. sulphureus*, *A. luchuensis*, *Candida albicans* *F.solani*, *Phythium sp* *Rhizopus sp*, *R.oryzae* and bacterial species are *S.lactis*, *Enterobacter sp*, *E. aerogenes* and *Pseudomonas sp*, *P.vulgaris*, *B. subtilis* *Staphylococcus sp*. In S2 site fungal organisms are *Aspergillus niger*, *A.itaconicus* *A.nidulans* *A.oryzae*, *Mucor sp*, *Fusarium oxysporium*, *Verticillium sp*, *Pythium*, *Helmithosporium* and bacterial species are *Micrococcus sp*, *Micrococcus sp* *Bacillus sp*, *Micrococcus sp* and In S3 site fungal organisms are *A.itaconicus*, *pencillium chrysogenum*, *R.nigricans*, *Candida albicans*, *Fusarium oxysporium* *Trichoderma viridae*, *Helmithosporium*, *R.oryzae* and bacterial species are *Azotobacter Bacillus sp*, *B. subtilis*, *Enterobacter sp*, *E. aerogenes* *E.coli*, *S.lactis* *P.aeruginosa* are dominants (Table 2,3).

Table - 2 Details of the bacteria isolated from soil

S.No	Bactreial Species	Monsoon			Postmonsoon			Summer			Premonsoon		
		S1	S2	S3	S1	S2	S3	S1	S2	S3	S1	S2	S3
1	<i>B. cerus</i>	-	+	+	-	-	+	+	-	-	+	-	+
2	<i>B. subtilis</i>	-	-	+	+	-	+	+	-	+	+	+	-
3	<i>B. circulans</i>	-	+	-	+	-	+	-	+	+	-	-	-
4	<i>B. megaterium</i>	+	+	-	+	-	+	-	+	-	+	-	+
5	<i>B. coagulans</i>	+	+	+	-	+	-	+	+	+	-	+	+
6	<i>B. mucoids</i>	-	+	-	+	-	+	+	-	+	-	+	+
7	<i>B. licheniformis</i>	-	+	-	+	+	-	+	-	+	+	+	-
8	<i>S. aureus</i>	+	+	-	+	-	+	+	+	-	+	+	-
9	<i>S. lactis</i>	+	-	-	+	-	+	-	+	-	+	-	+
10	<i>S. phyogens</i>	+	-	+	-	+	+	-	-	+	+	-	-
11	<i>E. aerogenes</i>	-	-	+	-	-	+	+	-	+	-	+	-
12	<i>E. coli</i>	+	-	-	-	+	-	-	-	+	-	+	+
13	<i>C. pyogens</i>	+	+	-	+	+	-	+	-	+	-	+	-
14	<i>P.aeruginosa</i>	+	+	-	-	-	+	+	-	+	-	+	-
15	<i>p.vulgaris</i>	+	+	-	+	-	+	+	+	-	+	-	+
16	<i>M.luteus</i>	-	+	+	-	+	+	+	+	-	+	-	-
17	<i>Azotobacter</i>	-	+	-	+	-	+	-	+	+	-	-	+
18	<i>Azospirillum</i>	+	+	-	+	-	+	+	-	+	-	-	+
19	<i>S. areus</i>	+	+	-	+	-	+	-	+	-	+	+	+
20	<i>Enterococcus</i>	-	+	-	+	-	+	+	-	+	-	+	+
21	<i>Vibrio sp</i>	-	-	+	-	-	+	+	-	+	-	+	+

S1 -Kumbakonam, S2-Krisnapuram, S3- Cholapuram (+ Present, - Absent)

Maximum number of fungi and bacteria were isolated from monsoon season, minimum number of fungi and bacteria isolated in postmonsoon season. Majority of soil fungi belongs to Ascomycota and Deutromycota.

Table - 3 Details of the fungi isolated from soil

S.No	Species	Monsoon			Postmonsoon			Summer			premonsoon		
		S1	S2	S3	S1	S2	S3	S1	S2	S3	S1	S2	S3
1	<i>Aspergillus flavus</i>	+	-	+	+	+	+	-	+	+	+	+	+
2	<i>A.oryzae</i>	-	+	+	-	+	-	+	-	+	-	+	+
3	<i>A.niger</i>	+	+	+	+	+	+	+	+	+	+	+	+
4	<i>A.fumigatus</i>	+	-	-	+	+	+	-	+	-	+	-	+
5	<i>A.terrus</i>	+	+	-	+	-	-	+	+	-	+	-	+
6	<i>A.nidulans</i>	+	-	+	-	+	+	-	-	-	+	+	-
7	<i>A.vesicular</i>	-	+	+	-	+	+	-	-	-	+	-	+
8	<i>A.itaconicus</i>	-	-	+	+	-	+	+	-	+	+	+	-
9	<i>A. luchuensis</i>	-	+	-	+	-	-	+	-	+	+	+	-
10	<i>A. sulphurous</i>	+	+	-	-	+	+	-	+	-	+	-	-
11	<i>Cladosporium sp</i>	+	-	+	-	-	+	-	-	-	+	-	-
12	<i>C. herbarum</i>	-	+	-	-	-	+	+	-	-	+	-	+
13	<i>Cunninghamella sp</i>	-	+	-	+	+	-	+	-	+	+	+	-
14	<i>Candida albicans</i>	+	+	-	+	-	-	+	+	+	-	+	-
15	<i>Fusarium oxysporium</i>	+	-	-	+	-	+	-	+	-	+	-	+
16	<i>F. solani</i>	+	-	+	-	+	+	-	-	-	+	+	-
17	<i>F. semitectum</i>	-	-	+	-	-	+	+	-	+	-	+	-
18	<i>Helmithosporium</i>	+	-	-	-	-	+	-	-	-	-	+	+
19	<i>Mucor sp</i>	+	+	-	+	+	-	+	-	+	-	+	-
20	<i>Rhizopus stolonifer</i>	+	+	-	-	-	+	+	-	+	-	+	-
21	<i>R.oryzae</i>	+	+	-	+	-	+	+	+	-	+	-	+
24	<i>R.nigricans</i>	-	+	+	-	+	+	+	+	-	-	+	-
25	<i>Trichoderma viridae</i>	+	+	-	+	-	-	+	-	+	+	-	-
26	<i>Penicillium chrysogenum</i>	+	+	-	+	-	+	-	+	-	+	-	+
27	<i>P.bovis</i>	+	+	-	+	-	+	-	-	+	+	-	-
28	<i>Phythium sp</i>	+	-	-	-	+	+	-	-	-	-	-	+
29	<i>Verticillium sp</i>	-	-	+	-	-	-	+	+	-	+	-	-

S1 -Kumbakonam, S2-Krisnapuram, S3- Cholapuram (+ Present, - Absent)

Diversity was found to be higher in postmonsoon season than summer. The seasonal variation and (%) frequency of the bacterial and fungal flora were statistically analyzed (Table 7,8)

Our finding similar to this, [Senthil kumar et al., 2009](#) collected 15 soil samples from three different stations namely Koraiyar river head, Saradi, and Xavier munai along the Muthupet Mangroves in Tamilnadu and examined by dilution plating method of PDA medium to assess fungal diversity and the population diversity. Out of 22 species screened the *Aspergillus* and *Penicillium* were represented as dominant one of each. In the present study also species like *Aspergillus* and *Penicillium* were common to all sites.

Our work is supported by V. Manimegalai 2011, population density during Monsoon season, maximum, fungal species was recorded in 2009-2010. In our study report also highlighted that fungal population also high in monsoon season and similar that dominant species are *Aspergillus*, *penicillium* were also the same to that study.

Our finding similar to that [Kalaiselvi and Panneerselvam, 2011](#), seasonal variation of soil fungal population in Thanjavur district, Tamilnadu viz., Nadur, Orathanadu, Punnainallur and Tholkappiyar Square totally 30 different species belonging to Ascomycetes and Phycomycetes were isolated by using PDA medium. The dominant species were *Aspergillus niger*, *Cunninghamella sp.* followed by *Trichoderma viridae*. During rainy season maximum fungal count was recorded in sub soil layer.

Table -4 Physico-chemical Parameters of the soil

S.No	Name of the parameters	Monsoon			Postmonsoon			Summer			premonsoon		
		S ₁	S ₂	S ₃	S ₁	S ₂	S ₃	S ₁	S ₂	S ₃	S ₁	S ₂	S ₃
1	PH	8.21	7.57	7.31	7.96	7.06	7.91	7.82	7.72	7.72	7.76	7.28	7.41
2	Moisture (%)	60	59	63	56	67	72	34	39	36	48	57	55
3	Temperature(°C)	38	36	39	42	45	47	59	62	67	35	36	29
4	carbon(%)	0.42	0.92	0.49	0.46	0.25	0.74	0.28	0.48	0.14	0.45	1.85	0.82
5	Nitrogen(Kg/ac)	87.6	89.3	81.1	90.28	89.5	73.2	87.5	87.9	90.6	91.6	84.6	90.2
6	Potassium(kg/ac)	81	73.6	72.4	68.9	69.1	66.7	74.6	71.2	71.5	72.3	78.8	70
7	Phosphorus(kg/ac)	4.12	5.15	3.11	2.15	4.31	4.2	3.19	2.57	1.24	1.54	4.35	3.24
8	Magnesium(ppm)	10.5	10.1	8.7	8	8.2	8.5	9.5	8.8	10	7.5	9.7	9.3
9	Calcium(ppm)	8.4	9.9	8.1	7.6	7.1	7.3	8.8	8.6	9.5	9.4	9.5	9.6
10	Copper(ppm)	0.78	0.79	0.84	1.9	1.4	1.8	0.96	0.88	0.92	1.8	1.6	1.9
11	Iron(ppm)	2.34	2.55	2.65	3.3	3.6	2.25	4.7	4.6	4.9	2.5	2.8	2.6
12	Zinc(ppm)	0.87	0.78	1.51	0.67	0.81	0.74	1.8	2.7	2.5	0.7	1.5	2.3
13	Manganese(ppm)	2.31	3.5	2.8	2.1	1.9	2.4	3.5	3.8	3.5	2.2	1.7	2.8

S1 -Kumbakonam, S2-Krisnapuram, S3- Cholapuram

Our work similar to that, seasonal variation of soil fungal population in Thanjavur dist, Tamilnadu viz., Kumbakonam, Krishnapuram and Cholapuram surrounding totally 29 different species belonging to Ascomycetes. Our study monsoon season higher amount of fungal population recorded.

The present study was correlated to that Prince, (2012) that studies on soil mycoflora from the sugarcane field in Thanjavur District, Tamilnadu. About 50 different species belongs to Phycomycetes and Deuteromycetes were isolated. In our report highlighted that Ascomycetes and Deuteromycetes were isolated.

Table – 5 Details of the bacterial Colony forming units (CFU)

S.No	Species	Monsoon	Post monsoon	Summer	Pre monsoon
1	<i>B. cereus</i>	34	42	34	36
2	<i>B. subtilis</i>	46	38	27	44
3	<i>B. circulans</i>	42	36	23	39
4	<i>B. megaterium</i>	43	44	21	29
5	<i>B. coagulans</i>	55	39	32	32
6	<i>B. mucoidis</i>	40	29	37	45
7	<i>B. licheniformis</i>	34	32	25	48
8	<i>S. aureus</i>	55	45	23	52
9	<i>S. lactis</i>	57	49	33	68
10	<i>S. pyogenes</i>	48	38	28	64
11	<i>E. aerogenes</i>	52	48	39	68
12	<i>E. coli</i>	68	61	42	59
13	<i>C. pyogenes</i>	64	57	28	57
14	<i>P. aeruginosa</i>	68	55	29	68
15	<i>p. vulgaris</i>	59	61	25	34
16	<i>M. luteus</i>	57	45	33	27
17	<i>Azotobacter</i>	68	66	27	23
18	<i>Azospirillum</i>	65	69	29	21
19	<i>S. aureus</i>	60	47	33	32
20	<i>Enterococcus</i>	56	48	27	37
21	<i>Vibrio sp</i>	53	51	24	35

Similar to that Sukumaran, (2013) reported by characterization of bacterial diversity in marine sediment at different season in karankadu, bacterial diversity incidence was quite common in special and season fluctuation. The present study revealed that totally 18 bacterial species were recorded during Premonsoon season.

Our study was correlated to Uma maheshwari et al., 2013, from that totally 36 different species were isolated from seven taluks of Thiruvarur Dt. They are dependent on the nature of substrate

and temporal regions that form colonization. Here our study revealed that totally 18 different bacterial, and 29 fungi were isolated and identified from Kumbakonam Taluks of Thanjavur Districts.

The contribution of soil organisms was very significant in many soil functions such as supporting the growth of plants, absorbing, neutralizing and transforming compounds that might otherwise become pollutants in the environment. Some studies dealt with the influence of plant community and other attempted to examine seasonal trends on soil microorganisms.

Table- 6 Details of the fungal Colony forming units (CFU)

S.No	Species	Monsoon	Post monsoon	Summer	Pre Monsoon
1	<i>A. niger</i>	56	42	34	39
2	<i>A. flavus</i>	54	39	27	33
3	<i>A. terreus</i>	45	36	23	31
4	<i>A. Fumigatus</i>	38	44	21	41
5	<i>A. oryzae</i>	58	42	32	33
6	<i>A. nidulans</i>	48	29	37	34
7	<i>A. Vesicular</i>	34	41	25	21
8	<i>A. itaconicus</i>	35	45	23	28
9	<i>A. sulphurous</i>	48	49	33	40
10	<i>A. lughuensis</i>	32	38	28	64
11	<i>Cladosporium sp</i>	68	61	42	59
12	<i>C. herbarum</i>	64	57	28	57
13	<i>F. solani</i>	59	61	25	34
14	<i>F. oxysporium</i>	57	45	33	27
15	<i>F. semiectum</i>	68	66	27	23
16	<i>R. stolonifer</i>	60	47	33	32
17	<i>R. oryzae</i>	56	48	27	37
18	<i>R. nigricans</i>	53	51	24	35
19	<i>P. chrysogenum</i>	32	23	68	34
20	<i>P. bovis</i>	33	33	64	35
21	<i>P. expansum</i>	25	28	59	48
22	<i>C. albicans</i>	21	42	57	32
23	<i>T. viridae</i>	30	38	34	68
24	<i>T. harizonum</i>	27	36	27	64
25	<i>Phythium sp</i>	31	44	23	37
26	<i>Verticillium sp</i>	21	39	32	25
27	<i>Helmithosporiumsp</i>	20	29	37	23
29	<i>A.alternata</i>	22	32	35	33

Table -7 Details of frequency of bacteria in isolated sites

S.No	Genus	Species	Average	S1	S2	S3	Total	Frequency
1	<i>Bacillus sp</i>	<i>B. cereus</i>	34	9	12	13	34	11.5
		<i>B. subtilis</i>	46	8	10	9	27	9.18
		<i>B. circulans</i>	42	6	9	8	23	7.82
		<i>B. megaterium</i>	43	10	6	5	21	7.14
		<i>B. coagulase</i>	55	11	8	13	32	10.8
		<i>B. mucoides</i>	40	7	6	7	20	6.80
		<i>B. licheniformis</i>	34	8	8	9	25	8.501
		<i>Total</i>	294					
2	<i>Staphylococcus sp</i>	<i>S. aureus</i>	55	12	10	11	33	20.62
		<i>S. lactis</i>	57	9	11	8	28	17.54
		<i>S. phyogens</i>	48	7	8	6	21	13.12
3	<i>Enterobacter sp</i>	<i>E. aerogenes</i>	72	9	8	11	28	38.88
4	<i>Escherichia sp</i>	<i>E. coli</i>	78	10	12	7	29	37.17
5	<i>Clostridium sp</i>	<i>C. pyogens</i>	64	8	11	6	25	39.06
6	<i>Pseudomonas sp</i>	<i>P. aeruginosa</i>	68	11	9	13	33	48.52
7	<i>Proteus sp</i>	<i>p. vulgaris</i>	59	8	10	9	27	45.76
8	<i>Micrococcus sp</i>	<i>M. luteus</i>	57	5	7	6	18	31.56
9	<i>Azotobacter sp</i>	<i>Azotobacter</i>	68	10	12	11	33	48.52
10	<i>Azospirillum sp</i>	<i>Azospirillum</i>	65	8	11	8	27	41.53
11	<i>Streptococcus sp</i>	<i>S. aureus</i>	60	10	7	6	23	38.35
12	<i>Enterococcus sp</i>	<i>Enterococcus</i>	56	8	6	7	21	37.51
13	<i>Vibrio sp</i>	<i>Vibrio sp</i>	53	9	8	6	23	43.39

Table – 8 Details of frequency of mycoflora in the isolated sites

S.No	Genus	Species	Average	S1	S2	S3	Total	Frequency (%)
1	<i>Aspergillus sp</i>	<i>A. niger</i>	56	9	13	10	32	17.14
		<i>A. flavus</i>	54	7	10	16	33	17.36
		<i>A. terreus</i>	45	8	8	9	25	15.58
		<i>A. Fumigatus</i>	38	9	6	6	21	14.68
		<i>A. oryzae</i>	58	11	9	10	30	16.69
		<i>A. nidulans</i>	48	8	12	7	27	16.02
		<i>A. Vesicular</i>	34	10	14	7	31	16.91
		<i>A. itaconicus</i>	35	5	7	9	21	14.68
		<i>A. sulphurous</i>	48	6	8	6	20	14.46
				<i>A. lughuensis</i>	32	5	9	8
2	<i>Cladosporium sp</i>	<i>Cladosporium sp</i>	25	5	5	3	13	2.95
		<i>C. herbarum</i>	19	6	3	6	15	1.89
3	<i>Fusarium sp</i>	<i>F. solani</i>	65	11	10	12	23	13.60
		<i>F. oxysporium</i>	56	9	13	8	20	11.83
4	<i>Rhizopus sp</i>	<i>F. semitectum</i>	48	7	8	6	21	12.42
		<i>R. stolonifer</i>	169	7	8	6	21	12.42
		<i>R. oryzae</i>	67	9	16	14	39	20.72
5	<i>Penicillium sp</i>	<i>R. nigricans</i>	63	14	10	9	33	17.55
		<i>P. crysogenum</i>	58	12	11	8	31	16.40
6	<i>Candida sp</i>	<i>P. bovis</i>	68	11	9	13	33	18.71
		<i>P. expansum</i>	59	13	10	11	34	19.54
7	<i>Trichoderma sp</i>	<i>C. albicans</i>	47	8	7	6	21	12.06
		<i>T. viridae</i>	174	8	11	8	27	31.76
8	<i>Phythium sp</i>	<i>T. harzinum</i>	40	10	7	6	23	23.05
		<i>Phythium sp</i>	85	10	7	6	23	23.05
9	<i>Verticillium sp</i>	<i>Phythium sp</i>	56	8	6	9	23	14.01
10	<i>Helmithosporium sp</i>	<i>Verticillium sp</i>	53	9	8	11	28	15.84
		<i>Helmithosporium sp</i>	42	6	4	7	17	14.04

Table – 9 Cultural characteristics of bacteria

S.No	Organism	Gram stain	Cultural Characteristics	Motility	Indole	MR	VP	Citrate Utilization	Catalase	Urease
1	<i>E. aerogenes</i>	-	Abundant thick, white, glistening growth	Motile	-	-	+	+	+	-
2	<i>E. coli</i>	-	White, moist, glistening growth	Motile	-	-	+	+	+	-
3	<i>B. cereus</i>	+	Abundant, opaque, white waxy growth	Motile	-	-	+	+	+	-
4	<i>C. pyogens</i>	+	Gray- white, convex growth	Motile	-	-	+	+	+	-
5	<i>M. luteus</i>	+	Circular, entire, convex with regular edges	Non motile	-	+	-	-	+	-
6	<i>P. aeruginosa</i>	-	White large wringled growth	Motile	-	-	-	-	+	-
7	<i>S. aureus</i>	+	Smooth raised, glistening with circular growth	Non motile	-	+	-	-	+	-
8	<i>p. vulgaris</i>	-	Thin blue- gray, spreading growth	Non motile	+	-	-	+	+	-
9	<i>S. lactis</i>	+	Thin even growth	Non motile	-	+	-	-	+	-
10	<i>M. luteus</i>	+	Circular, entire, with regular edges	Non motile	-	+	-	-	+	-

Table – 9 Cultural characteristics of bacteria

11	<i>B.megaterium</i>	+	Abundant, opaque, white waxy growth	Motile	-	+	-	+	-
12	<i>B. coagulase</i>	+	White large wringled growth	Motile	-	-	+	+	-
13	<i>Azotobacter</i>	-	Thin blue- gray, spreading growth	Non motile	-	+	-	+	-
14	<i>Azospirillum</i>	-	gray, spreading growth	-	-	+	-	-	-
15	<i>B. subtilis</i>	+	Abundant thick growth	Motile	+	-	-	+	-
16	<i>S. phyogens</i>	+	Smooth raised growth	Non motile	-	-	+	+	+
17	<i>B. circulans</i>	-	Circular, entire, with regular edges	Motile	-	+	-	-	+

Table -10 Cultural characteristics on a PDA of the isolated fungi

S.No	Species	Upper surface			Lower surface	Observations
		Cultural Aspect	Density	Colour		
1	<i>Rhizopus stolonifer</i>	Effuse cotton	High	White at first become bluish black maturity	Idem to upper face	Rhizords rare sporangiophere
2	<i>Aspergillus flavus</i>	Effuse floccose	Medium	Conidial heads yellow to green	Idem to upper case	Hyphae, Septate with conidiophore
3	<i>A.oryzae</i>	Effuse globose	High	Orange to vinaceous or purple sclerotia	Idem to upper face	Hyphae, Septae with conidiophore
4	<i>Pencilium bovis</i>	Effuse floccose	Light	Grey green to brownish	Idem to upper face	Conidiophore (vertical of phialides)
5	<i>P. chrysogenum</i>	Effuse floccose	Light	Yellow to green	Idem to upper face	Conidiophore, compact vertical of phialides
6	<i>Trichoderma viride</i>	Effuse globose	Medium	Light green	Idem to upper face	Conidio elliptical and septae
7	<i>Fusarium sp</i>	Globose	Light	Grey colour	Idem to upper face	Conidia and septae
8	<i>Rhizopus sp</i>	Effuse cotton	Medium	Conidial heads yellow to green	Idem to lower face	Conidiophores
9	<i>Verticillium sp</i>	Thread like apperance	Light	Light to green White	Idem to lower face	Hyphae, Septae with conidiophore

CONCLUSION

The present study was planned to, Isolation and Identification of soil bacteria and fungi from Kumbakonam taluk of Thanjavur (Dt) Tamilnadu deals with diversity and distribution of bacterial and fungal population in an around soil. The physico – chemical parameters of such soils were recorded. Population of soil bacteria and fungi might also get affected by climate and resistance over extreme environmental condition.

Bacterial and fungal species are especially important components of biodiversity as major contributors to the maintenance of the earth's ecosystem, biosphere and biogeochemical cycle fungi perform unique and indispensable activities on which larger organism including human depend. The present study could be concluded that there is no uniformity in the diversity of population during season and their distribution pattern in different geographical regions. Several factors of salinity, origin, nature of substrata, pH and diversity bacteria and fungi. So it is obvious that a study based on biodiversity is a major challenging task as we try to predict the secret of nature.

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