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CODEN: IJRSFP (USA)

International Journal of Recent Scientific Research Vol. 8, Issue, 7, pp. 18427-18430, July, 2017 International Journal of Recent Scientific Re*r*earch

DOI: 10.24327/IJRSR

Research Article

ASSESSMENT OF CYANOBACTERIAL POPULATION IN THREE DIFFERENT WATER RESERVOIRS OF SIVAKASI

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DOI: http://dx.doi.org/10.24327/ijrsr.2017.0807.0506

ARTICLE INFO	ABSTRACT		
Article History: Received 20 th April, 2017 Received in revised form 29 th May, 2017 Accepted 30 th June, 2017 Published online 28 th July, 2017	Cyanobacteria live in a diverse range of environments with a number of features which contribute to their success. In the present study, the cyanobacterial population of three different ponds of Sivakasi (Virudhunagar district, Tamilnadu, South India) was studied. During the period September 2014 to October 2015, the survey revealed the presence of 59 species of cyanobacteria belonging to 14 genera and four families. The physico-chemical parameters like pH, temperature, carbonate, bicarbonate, nitrate, nitrite, ammonia, total phosphorus, inorganic phosphorus, calcium, magnesium, chloride and algal flora were studied in all the three water sources. In all the three pond systems, the		
Key Words:	species observed were Lyngbya, Oscillatoria, Phormidium, Microcystis, Synechococcus, Gloeocapsa, Synechocystis, chroococcus, Arthospira, Nostoc, Anabaena, Spirulina, Plectonema and		

Freshwater habitat, cyanobacterial population, identification of cyanobacteria, physico-chemical parameters.

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INTRODUCTION

Cyanobacteria, the oxygenic, photosynthetic prokaryotes are found in almost every aquatic and terrestrial environment. They show remarkable degree of morphologial and developmental diversity. Traditionally, their taxonomy is based upon morphologically diverse having unicellular, colonial and filamentous form (Whitton and Potts, 2000). They play a vital role in the various biogeochemical cycles such as nitrogen, carbon, phosphorus and oxygen. For a long time, the economic importance of cyanobacteria was limited to their use as biofertilizer in agriculture but during the last few decades they have been recognized for their high potential in a wide variety of biotechnological application. (Thajuddin and Subramanian, 2005; Spolaore et al, 2006). Cyanobacteria are commonly found in polluted and non-polluted waters and due to this behavior they are generally considered useful to determine the quality of water at very low cost. They are very suitable organisms for the determination of the impact of toxic substances on the food chain and also have consequence on the higher level. Clean water would support a great diversity of organisms, with one or few dominant forms (Trainer, 1984).

Few reports have been available on cyanobacteria flora from different regions of India (Desikachary, 1959; Prasad and saxena, 1980; Tararand Bodkhe, 1997, Mahadevand Hosmane, 2004; Parikh, et al, 2006; Gupta, et al., 2006; Sankaran, 2006; Chaudhary and Kumar Mukesh, 2006; Saha, et al., 2007; Kumar Mukesh, 2010; Kumar Mukesh, et al., 2011). Some algal infestations have been reported from Kanpur by various workers in time and space (Ahked, 1973; Shukla, 1983; Tripathi and Pandey, 1989; Tiwari, et al., 2001; TiwariandShukla, 2007, Rishi and Awasthi, 2012). During the past few decades research on cyanobacteria were mostly academic oriented and now proved as potential candidates for biotechnological utilization (Richmond, 1990; much Sundararaman and Sekar, 2001; Thajuddin and Subramanian, 2005). Cyanobacteria have been an unexplored arena for biological researchers at large. Mostly the microbial analysis was done taking the bacterial and fungal strains into account leaving behind the mostly notable autotrophic microbial strains in the name of algae. Cyanobacteria strains which do possess the character of nitrogen fixation add up to value addition of these microbial strains mainly in the field of man culture, food,

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feed, fuel, fertilizer, medicine and combating pollution (Muthukumar *et al.* 2007)

MATERIALS AND METHODS

Sampling

Algal samples were collected from three different ponds in and around Sivakasi. Algal species inhabiting these water bodies were collected in the sterilized polythene bags and wide mouthed bottles. The samples were brought to the laboratory for isolation, purification and identification purposes. Collected samples were examined to identify the algal species. Isolation and purification of algal strains was done by repeated sub culturing on solidified and in liquid medium (BG11) by dilution and pour plate technique. The samples were then placed in an incubator at 28 ± 1^{0} C, with 16h light/dark cycle. Light was provided by a single 58 watt fluorescent tube (luminex, cool white) at 25μ molQuanta m⁻² s-1 for 18 days. Each sample was observed under Olympus light microscope. The different environmental parameters data prevailing on each of the sampling site was studied.

 Table I Physiochemical parameters of water samples in three water stations

Sl.NO	PARAMETER	POND I	POND II	POND III
1	pН	7.5	6.5	7.5
2	Temperature	30	31	30
3	Carbonate	40	30	30
4	Bicarbonate	90	110	120
5	BOD	120	110	100
6	COD	632	744	624
7	DO	3.7	3.1	3.35
8	Nitrate	59	66	62
9	Nitrite	34	32	34
10	Ammonia	276	261	284
11	Total phosphorus	88	66	102
12	Inorganic phosphate	32	36	40
13	Organic	56	30	62
14	Calcium	50.46	54.66	46.25
15	Magnesium	19.40	33.02	30.19
16	Chloride	159	119	114

RESULTS AND DISCUSSION

In the present Study, the cyanobacteria community of three different ponds of Sivakasi was studied. Totally 59 species of cyanobacteria belonging to four families, 13 genera have been identified from different sampling stations during the period from September 2014 to October 2015 (Table 2). Among them, pond II recorded the maximum number of species (46) followed by pond I (38) and pond III (28). Among the ponds, heterocystous forms were identified in pond I and II and were not recorded in pond III. Heterocystous cyanobacteria such as Nostoc calcicola was recorded in pond I and II. On the other hand Nostoc linckia. Anabaena aphanizomenoides were recorded only in pond II, while Nostoc calcicola recorded in pond I and II (Table 2). In total, 20 species of cyanobacteria were recorded in common to all the ponds analyzed. Of them, Oscillatoria with 15 species was the dominant genus, followed by Phormidium (11), Lyngbya(10), Microsystis (3) Synechococcus (2) and Synechocystis with single species (Table 2). The rich growth of cyanobacteria is believe to be due to favorable content of oxidisable organic matter and less dissolved oxygen (Table I) an observation which supports Vijayakumar et al., (2007) Gomathi et al., (2011) and Mathumathi and Vijayakumar (2013).

Table II Population density of cyanobacteria in three						
different pond system						

		Pond-I	Pond-II	Pond-III
Sl.No	Cyanobacterialspecies) (percentage)	
1.	Synechocystis aquatilis	33	n.d	41
2.	Synechococcus elongatus	66	33	25
3.	Chroococcus minor	33	58	n.d
4.	Chroococcus sps.	66	68	58
5.	Microcystis flos aquae	75	58	66
6.	Microcystis robusta	41	n.d	75
7.	Microcystis aeruginosa	100	100	83
8. 9.	Arthrospira platensis Spirulina platensis	75 n.d	66 91	n.d n.d
9. 10.	1 1	25	n.d	n.d
10.	Spirulina sps. Oscillatoria curviceps	23 50	66	25
12.	Oscillatoria earlei	100	91	100
12.	Oscillatoria late-virens	91	100	83
14.	Oscillatoria limentica	n.d	50	75
15.	Oscillatoria princeps	n.d	75	66
16.	Oscillatoria pseudogerminata		91	91
17.	Oscillatoria rubescens	41	n.d	50
18.	Oscillatoria salina	100	83	16
19.	Oscillatoria subbrevis	83	75	66
20.	Oscillatoria willie	91	66	41
21.	Phormodium ambiguum	75	50	66
22.	Phormodium corium	91	83	100
23.	Phormodium tenue	83	75	75
24.	Phormodium uncinatum	n.d	25	16
25.	Phormodium laminosum	n.d	8	n.d
26.	Phormodium anomala	25	n.d	n.d
27.	Oscillatoria tereberiformis	91	100	83
28.	Oscillatoria brevis	75	50	41
29.	Oscillatoria animalis	n.d	75	n.d
30.	Oscillatoria chlorine	25	8	16
31.	Lyngbya aestuarri	50	75	8
32.	Lyngbya borgerti	25	8	16
33.	Gloeocapsa polydermatica	75	50	66
34.	Lyngbya confervoides	91	83	75
35.	Lyngbya martensiana	66	n.d	41
36.	Lyngbya majuscule	25	50	16
137.	Lyngbya putealis	16	n.d	8
₂ 38.	Lyngbya spiralis	n.d	50	66
39.	Lyngbyatrunicola	66	75	83
40. 41.	Nostoc linckia	n.d 66	100 91	n.d
41. 42.	Nostoc calcicola Anabaena aphanizomenoides	00 n.d	41	n.d n.d
42.	Plectonema radiosum	n.d	41	n.d
44.	Chroococcus minutus	25	n.d	n.d
45.	Chroococcus schizodermatica		n.d	n.d
46.	Chroococcus minor	16	n.d	n.d
47.	Phormidium africanum	50	n.d	n.d
48.	Phormidium sps.	66	n.d	n.d
49.	Oscilatoria limosa	n.d	n.d	n.d
50.	Phormidium foveolorum	n.d	41	n.d
51.	Phormidium mucicole	n.d	50	n.d
52.	Arthospira tenuis	n.d	66	n.d
53.	Synechococcus cedroren	n.d	50	n.d
54.	Phormidium jenkelianum	n.d	41	n.d
55.	Arthospira massartii	n.d	50	n.d
56.	Chroococcus gomontii	n.d	66	n.d
57.	Phormidium fragile	60	75	80
58.	Lyngbya gracilis	n.d	50	n.d
59.	Aphanocapsa sps.	n.d	81	n.d

The study reveals that calcium is one of the important minerals which supports the growth of Cyanobacterial population. Favouring the growth of cyanobacteria is not only by calciumbut also high amount oxidizable organic matter, traces of dissolved oxygen, considerable amount nitrate and phosphates also Hamed *et al.* 2008, Senthil *et al.* 2012, Vijayakumar, 2012 and Mathumathy and Vijayakumar, 2013 Vijakumar *et al.*, 2005 Gomathi *et al.* 2011 reported that

cyanophyceae grow luxuriantly with great variety abundance in ponds having rich calcium Muthukumar *et al.* (2007) and Vijayakumar and Manoharan (2012) reported that high values of BOD, COD, phosphates and nitrate with very low DO favored the growth of algae. Their findings were supported by Jaeyachitra *et al.* (2013) and Vinoth Rishi and Awasthi (2015). The present study showed considerable amount nitrate and phosphate with increased level of BOD, COD along with very low DO level. However, Senthil *et al.* (2012) observed that, correlation between abundance of planktonic cyanobacteria and concentration of dissolved oxygen could be due to the depletion of oxygen.

In the present study, heterocyst us cyanobacteria Anabaena aphanizomenoides, Nostoc linckia, Nostoc calcicola were observed (Table 2). Hamed et al (2008) Gomathi et al (2011) Mathumathy and Vijayakumar (2012) reported that Calothrix, Nostoc carneum and Anabaena species in fresh water lakes. Gurstafsom et al (2008) and Gomathy et al (2011) reported that the genus Oscillatoria has been found tobe very tolerant and frequently inhabit the polluted waters. Similarly, Mathumathy Vijayakumar (2012) and Jeyachithra et al (2013) reported Oscillatoria, Phormidium were the most dominant genera in lakes and ponds respectively. The present study reveals the presence of Oscillatoria. Phormidium along with Lyngbya in all the ponds studied (Table). Senthil et al. (2012) emphasized the use of algae as reliable indicators of pollution. Certain members of Cyanobacteria are tolerant to organic pollution and resist environmental stress caused pollutant such species can be used as indicators particular habitat (Vijayakumar, 2012). In the present study, pond I with 13 species followed by pond II with 19 species and pond III with11 species each (Table II), the percentage values of which were 75and above should be considered as indicators of the respective ponds are reported by Gomathy et al. (2011). Of the indicator species observed in different ponds, Microcystis aerunginosa, Oscillatoria earlei, Phormidium corium were found to be more than 80% representation in all the ponds and thus considered as an indicators of the algal population. From the foregoing discussion, it is concluded that the physicochemical characters together with biological monitoring provided converging lines of evidence for evaluation of polluted habitat in this case as in some other studies (Murugasen, 2005; Gomathi, 2011).

CONCLUSION

The present study, undertaken to analyze the presence of cyanobacterial population revealed that no significant variation in the occurence and distribution of cyanobacteria which would be correlated to the physico-chemical parameters of water. The cyanobacterial species investigated in the present study are highly recommended for beneficial bioremediation applications for *in-situ* and off-site removal of pollutants.

Acknowledgement

The author would like to express her sincere thanks to the principal and management for providing lab facilities and also thank Guide and Co- guide for their constant support and encouragement in this work.

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How to cite this article:

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Maheswari S et al.2017, Assessment of Cyanobacterial Population in three Different Water Reservoirs of Sivakasi. Int J Recent Sci Res. 8(7), pp. 18427-18430. DOI: http://dx.doi.org/10.24327/ijrsr.2017.0807.0506
