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Research Article

A REPORT ON THE PREDATORY BEHAVIOUR OF STINK BUG EOCANTHECONA FURCELLATA WOLFF (HEMIPTERA: PENTATOMIDAE) ON THE EGGS AND GRUBS OF HENOSEPILACHNA VIGINTIOCTOPUNCTATA

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ABSTRACT

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Key Words:

Biological control, E. furcellata, H. vigintioctopunctata, S. melongena.

Henosepilachna vigintioctopunctata (Coleoptera: Coccinellidae) is a threat to commercial plantations of *Solanum melongena* species in Asia and there are no effective strategies for its control. We report here for the first time, the occurrence of the predatory stink bug *Eocanthecona furcellata* (Hemiptera: Pentatomidae) nymphs and adults effectively feeding on *H. vigintioctopunctata* eggs and grubs. *Eocanthecona furcellata* is a native generalist predator that is easily reared in the laboratory and may be utilized as a biological control agent. Thus, *E. furcellata* can be used as a natural enemy in biological control programs targeting the pest on brinjal.

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INTRODUCTION

Henosepilachna vigintioctopunctata F. is a polyphagous plant feeder and most destructive pest of many cultivated (brinjal, tomato, potato, tobacco, melon, cucumber, gourds, and pumpkin) and wild crops (Ahmad et al, 2001; Rath, 2005). It can be easily identified by its 28 spots on its dorsal body. It is also known as Hadda beetle or Ladybird beetle. This beetle is fairly common and cause damage to the Solanaceous and Cucurbitaceous crops (Khan et al, 2000). The peak infestation period of this pest varies from region to region. Both adult beetles and grubs feed on the epidermal tissue of the leaves by scrapping on the leaf surface. This results in the leaves becoming dry and eventually shed. Sometimes it is called leaf scrapping Coccinellid beetle (Imura and Ninomiya, 1978). In severe cases, even the calyx of the fruits may also be infested (Varma and Anandhi, 2008). The growth and development of the plants are greatly hampered and the yield is markedly reduced by the attack of this pest. (Alam, 1969). The increasing pest status and abundance of hadda beetle has raised a number of questions regarding the factors responsible for its development, its natural enemy complex and management under natural conditions.

Biological control is recognized as one of the best alternatives to the use of chemical insecticides for controlling insect pests. Pest control with natural enemies is growing in popularity and heteropteran predators are important agents of biological control in integrated pest management programs (Molina et al, 1997; Lemos et al, 2003). Thus, biological control, such as the introduction or enhancement of populations of predators, parasitoids and pathogens, is important (Silva, 2000). Heteropteran predators are important biological control agents on leaf worms (De Clercq et al, 2003; Lemos, 2003), beet armyworms (De Clercq and Degheele, 1994), Colorado potato beetles (Biever and Chauvin, 1992; Hough-Goldstein and McPherson, 1996; Tipping et al, 1999; Westich and Hough-Goldstein, 2001), and southern green stinkbugs, respectively (De Clercq et al, 2002) on soybean caterpillars (Marston et al, 1978).

The most common pest management activity that negatively impacts beneficial organisms in agroecosystem is pesticide application. As a result, modifications of pesticide use practices are the most commonly implemented form of conservation biological control (Ruberson *et al*, 1998), and have long been considered an important component of integrated pest management programs (Stern *et al*, 1959; Debach, 1964; Newsom and Brazzel, 1968). Pesticide use can be modified to

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favour natural enemies in a variety of ways, including treating only when economic thresholds dictate, use of active ingredients and formulations that are selectively less toxic to natural enemies, use of the lowest effective rates of pesticides, and temporal and spatial separation of natural enemies and pesticides (Hull and Beers, 1985; Poehling, 1989; Ruberson *et al*, 1998). Decisions regarding pesticide use for insect pests in IPM programs are typically based on sampling pest populations to determine if they have reached economic threshold levels (Pedigo, 1989).

E. furcellata has been reported from Southeast Asia, Japan, India, and Taiwan, and has been preying on Lepidopteran, Coleopteran and Heteropteran insects (Ahmad *et al*, 1996; Chu 1975; Chang, 2002; Jakhmola 1983; Prasad *et al*, 1983). However, there is no publication available on the predation efficacy of *Eocanthecona furcellata* with regard to *H. vigintioctopunctata* larvae feeding on any host plants. From this study, we observed that *E. furcellata* reached the adult stage when it fed only on *H. vigintioctopunctata* eggs and grubs, which suggest that could keep this prey population in control.

MATERIALS AND METHODS

Third nymphal instars of predatory stink bug Eocanthecona furcellata were originally collected from the brinjal field at MCC Farm, Chennai, Tamilnadu, India. The collected nymphal instars were carefully introduced into an insect cage. Meanwhile eggs, second and third grub stages of Henosepilachna vigintioctopunctata were also collected from the brinjal field and were served as feed for the nymphal instars of Eocanthecona furcellata every morning and themorphological changes from third nymphal in star into adult were observed in the laboratory condition too. The experiment was conducted under the room temperature (32°C). Each stages of the predatory bug were easily identified by its exuviae, and also the colour variation on its dorsal surface.

Species Identification

The identified predator species was confirmed by determining them using the keys and descriptions (Distant, 1902; Ananthi Rachel and Gladstone, 1993) as *Eocanthecona furcellata*, (Wolff). 16mm long, 8mm broad, elongately ovate, brownish, ochraceous, densely darkly punctate, punctures bronzy.

Head: Almost as long as broad across the eyes; frontal and loral lobes almost at the same level apically, loral lobes expanded and rounded; two piceous longitudinal lines, the inner one terminating at the middle of the frons and the outer one extending up to the vertex; antennae five jointed, basal joint not passing head, apical two segments with basal stramineous rings; eyes dark brown; ocelli brick red, close to the eyes; rostrum reaching the metasternum; apical two joints piceous; basal two joints ochraceous, slightly incrassated.

Thorax: Pronotum with broken transverse brownish fascia anteriorly; disc darkly coarsely punctate, marginally sinuate; anterolateral angles minutely tuberculate; lateral margins of anterior lobe minutely dentate; posterolateral angles of the disc spinously produced, spine bifid, anterior ramus longer; piceous; scutellum elongately triangular, bronzy black at base, basilateral angles with lavigate spot, posteriorly tapering as

lavigate spatulate tip, reaching the sixth abdominal segment; corium with bronzy black suffusions; membrane transparent with elongate brown marking basally and medially, extending far beyond the abdomen; legs reddish ochraceous; junctions of joints piceous; thoracic sternites, and connexivum coarsely punctate with large subquadrate bronzy black spots; marginal spots to abdomen, a central black spot at the sternite of genital segment.

RESULT AND DISCUSSION

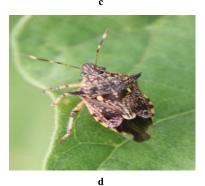
From this it was observed that, the eggs and grubs of H. vigintioctopunctata is edible and a natural prey for this predator bug (Table 1, Fig 1). During the study, when the grubs were introduced into the rearing cage of E. furcellata nymphs, the process of feeding started by sucking the haemolymph of grubs continuously and finally skeletonizing them. In case of prey's egg, they made a hole on the top of eggs surface and made them into empty shell. Yasuda, 1997 and 2000 proved that E. furcellata located their prey by chemical cues emanating from their prey. According to (Khin 2008) the predatory bug E. *furcellata* may detect the prey by vibrations caused by feeding. Khin 2008, have studied that the predation efficacy of E. furcellata was tested with American Bollworm from four different host plants (Cabbage, cotton, chickpea and tomato plants). The majority of E. furcellata (30-60%) directly approached towards cotton plants sucking on H. armigera larvae. Most probably, E. furcellata responded to (E)- phytol, which is produced by larvae when feeding on the chlorophyll in their food plants. E. furcellata prefers to feed on larvae fed with a chlorophyll-rich diet rather than with a chlorophyll-poor diet (Yasuda, 1997, 1998a, 1998b).

Table 1 Normal life cycle of *E. furcellataon H. vigintioctopunctata*

S.No	Stage	Longevity	Given as prey
1	3 rd nymphal instar	3-4 days	Grubs
2	4 th nymphal instar	3-4 days	Grubs & egg batches
3	5 th nymphal instar	5-6 days	Grubs & egg batches
4	Adult	20-25 days	Grubs, egg batches, semi looper caterpillars, larvae of <i>Leucinodes orbonalis</i>

















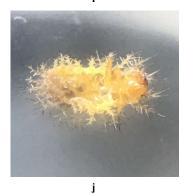


Fig 1 A- Third nymphal instar of *E. furcellata*, B- Fourth nymphal instar, C- Fifth nymphal instar, D- Adult of *E. furcellata*, E- Exuviae, F- Normal egg of *H. vigintioctopunctata*, G- nymphal instar feeds egg batches, Hempty egg batches, I- nymphal instars feeds grub of *H. vigintioctopunctata*, J- skeletonized body of grub.

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