

Available Online at http://www.recentscientific.com

**CODEN: IJRSFP (USA)** 

International Journal of Recent Scientific Research Vol. 9, Issue, 1(C), pp. 23023-23027, January, 2018 International Journal of **Recent Scientific Re**rearch

DOI: 10.24327/IJRSR

# **Research Article**

## DO DROSOPHILA LARVAE PREFER LIGHT OR DARK?-AN IMPROVISED INVESTIGATORY **PROJECT FOR HIGH SCHOOL STUDENTS TO GO BEYOND TEXT BOOK**

Nagaraj G\*

Department of Education in Science and Mathematics, Regional Institute of Education, (National Council of Educational Research and Training), Mysuru, Karnataka, India

DOI: http://dx.doi.org/10.24327/ijrsr.2018.0901.1383

ARTICLE INFO	ABSTRACT						
Article History: Received 15 <sup>th</sup> October, 2017 Received in revised form 25 <sup>th</sup> November, 2017 Accepted 23 <sup>rd</sup> December, 2017 Published online 28 <sup>th</sup> January, 2018	Present study aims to explore the phototaxic behavior of larvae of three <i>Drosophila melanogaster</i> strains (namely Oregon K, sepia eye and white eye) with an improvised low-cost setup, which in turn may be a useful project for the school students of IX - XII classes. Simple light-dark preference test was carried out using a pair of glass Petri dishes with dark and light area, small cupboard and LED torch emitting 1000 lux light. Number of larvae in light and dark area was captured using a smart phone and their percentage was calculated. Results showed that all the three strains of larvae were negatively phototaxic and normal O.K larvae were more (84%) sensitive to light than the						

#### Key Words:

Phototaxis, practical classes, mutants, transition, rava-jaggery' medium.

were more (84%) s mutant sepia (70%) and white (71%) eyed larvae. There may exist a relation between phototaxis response and eye color. Further, as the pre and in-service teachers opined, this study can be an effective project for the higher secondary students for active learning by doing and to go beyond text book.

Copyright © Nagaraj G, 2018, this is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

## **INTRODUCTION**

Appropriate preference for light or dark can be crucial for an animal's survival (Gong et al., 2010). Studies found two types of photo-receptor neurons (PRs) expressed by either Rhodopsin5 and Rhodopsin6 in the larval eye (Salcedo et al., 1999, Sprecher et al., 2007; Keene et al., 2011; Hassan et al., 2005) and Humber and Sprecher (2017) found that only the Rhodopsin5 subtype is essential for light avoidance. Gong et al. (2010) reported that two pairs of isomorphic neurons in the central brain switch the larval light preference. Further, Xiang found that light-avoidance-mediatingal. (2010)et photoreceptors tile the larval body wall. Carl Friendrich and Thomas (2017) mappd the larval eve's mind to understand the neural circuitary of larval vison. Further, Zhao et al. (2017) stated that turns with multiple and single head cast mediate Drosophila larval light avoidance. Studies also stated that, visual pathways which mediate Drosophila larval light avoidance and circadian clock entrainment are distinct (Keene et al., 2011) and are developmentally related (Hassan et al., 2005). Rodriguez and Campos (2009) reported that inactivation of DOPA decarboxylase neurons increases the response to light throughout larval development and Mapel et al. (2002) found the strong role of norpA-encoded phospholipase C signaling

along with RH5 and Rh6 in both larval and adult extra-retinal circadian photoreception. Studies as well compared the larval and adult photoreception mechanisms and stated that larval and adult photoreceptors use different mechanisms to specify the same Rhodopsin fates (Sprecher et al., 2007); and same transcription factors regulate diverse aspects of larval and adult photoreceptor development at different stages and in a contextdependent manner (Mishra et al., 2013). Furthermore, Rodriguez and Campos (2009) stated that Drosophila larval photobehavior can be used to study the control of locomotion. Practical classes bring the theory in action to verify Science concepts and provide empirical evidences. Not only that, they develop problem solving and reasoning ability, enhance mastering the subject, provide the scope for learning by doing, inculcates practical skills, trains in scientific methods and cultivates scientific temper (NCERT, 2006). Studies proved that, students learning with lots of experiments understand the concept well and get high marks in exams (Emerson and Taylor, 2004; Ball et al., 2006). Correspondingly, Frank (1997) finds that, compared to students in a control class, students' homework scores increase when they participate in an experiment related to the homework topic.

<sup>\*</sup>Corresponding author: Nagaraj G

Department of Education in Science and Mathematics, Regional Institute of Education, (National Council of Educational Research and Training), Mysuru, Karnataka, India

Teachers sometimes could not conduct effective practical classes because of non-availability of proper equipments or space especially in rural schools. In such situation low cost, improvised experiments which can be performed even at home can be very useful. In this context, present study aims to study the phototaxis behavior (light/ dark preference) of larvae of *Drosophila melanogaster* with an improvised low-cost setup, which in turn may be a useful experiment/ project for the school students of IX - XII class.

### **MATERIALS AND METHODS**

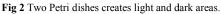
Studies in the past used various highfy material and methods for deeper understanding of phototaxis. For example, high resolution computer based tracking analysis (Keene *et al.*, 2013; Gershow *et al.*, 2012; Hernandez-Nunez *et al.*, 2015) to quantify sensory motor structure of larvae. However, present study adapts/ improvises simple light-dark preference test as carried out by earlier studies (Gong, 2009; Gong and Gong, 2012; Farca Luna, 2013). A small cupboard (as dark box) having the dimension of 100x50x30 cm was taken and a white porcelain tile was kept at it's floor as shown in fig. 1.



Fig 1 Improvised experimental setup.

An LED torch was made hanging from the roof of the box and the torch emitted white light broad band spectra ranging from 450-600nm. The light falls on the larvae from 75cm height with 1000 lux intensity. Students can use even old carton boxes and small torch light. A pair of glass Petri dishes having the diameter of 15cm was taken and divided into four quarters. In each dish, opposite quarters and its' side wall was pasted with black paper as shown in fig. 2.





Three pure strains of *Drosophila melanogaster* viz. normal red eye (Oregon K) and mutants sepia eye and white eye flies were

brought from the University of Mysore. These flies were cultured in glass bottle at room temperature with 12hr dark and 12hr light period in 'rava-jaggery' medium (Nagaraj, 2016). School students can collect the wild flies by keeping smashed banana in a bottle near the window. They can get larvae by rearing the flies in 'rava-jaggery' medium.

Mostly III instar larvae of O.K strain counting 10 numbers were collected from the side walls of the bottle using a brush. They were rinsed with Ringer's solution to remove the adhering food particles and kept in a plain Petri dish for 20 min for acclimatization. Meanwhile, one of the black paper pasted-Petri dishes was kept on the floor of the box. Then those larvae were transferred to the centre of this Petri dish and covered using another paper pasted Petri dish. The pasted black papers of two Petri dishes (upper and lower) were co-insides each other in order to create dark and light area (fig. 2) in the Petri dish. The torch light was ON and the door was closed in order to prevent the influence of external light, so that only the source light (torch light) effects the behavior of larvae. For every 5 minutes the door was opened very little and the position or number of larvae light and dark area was captured using a smart phone (without flash light) for about 2hrs. From the captured pictures the number and percentage of larvae in light and dark area was calculated and tabulated. Similar assay was performed for sepia and white eyed larvae in the next day.

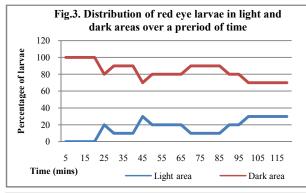
### **RESULTS AND DISCUSSIONS**

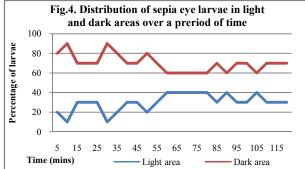
It is observed from the table 1 that, all the three strains of larvae showed negative phototaxis response. In support, to the present observation, other studies also found that the larvae are negatively photactic (Humberg and Sprecher, 2017; Sawin-McCormack et al., 1995). Further, the average percentages of larvae in dark area shows that, among the three strains the normal O.K larvae are more (84%) sensitive (negatively) to light than the mutant sepia (70%) and white (71%) eyed larvae. Likewise, Sawin-McCormack et al., (1995) observed that glmutant larva failed to respond to light during the foraging state and opined that this is likely due to lack of larval photoreceptors. Gong (2009) observed the preference of light to darkness by tim(01) larvae in the immediate light/ dark boundary passing test. Similarly, studies were also noted that,  $norpA^{P41}$ ;  $cry^b$  double mutants were more affected than simple mutants in their entrainment to light-dark cycles (Emery et al., 2000; Stanewsky et al., 1998). In addition, average percentages also show that both mutants are equally sensitive (negatively) to light.

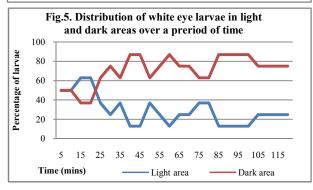
Moreover, the pattern of response which may be noted from the fig. 3-5 shows that, as the time progress red and sepia eyed larvae showed little inclination towards light (fig. 3 and 4), which may be due to acclimatisation / adjustment to light. On the contrary, white eyed flies initially moved towards light (not all) but, returned back to the dark as the time progress (fig. 5). It may be inferred from such response pattern that, there may exist a relation between phototaxis response and eye color (though proper eye was not developed). My previous study (Nagaraj, 2016) conducted with adult flies of same above three strain showed that, the flies are positively phototaxis and among them sepia eyed flies were fast in responding to light. Fascinatingly, studies also observed that, the larva in the earlier stage (I and II instar) was photo-negative (Mazzoni et al., 2005; Scantlebury et al., 2007; Yamanaka et al., 2013), whereas in the later stage (III instar) it became photo-neutral

Strain Time (mins)	Red eye (N=10)				Sepia eye (N=10)				White eye (N=8)			
	Light		Dark		Light		Dark		Light		Dark	
	Ν	%	Ν	%	Ν	%	Ν	%	Ν	%	Ν	%
5	0	0	10	100	2	20	8	80	4	50	4	50
10	0	0	10	100	1	10	9	90	4	50	4	50
15	0	0	10	100	3	30	7	70	5	63	3	37
20	0	0	10	100	3	30	7	70	5	63	3	31
25	2	20	8	80	3	30	7	70	3	37	5	63
30	1	10	9	90	1	10	9	90	2	25	6	7:
35	1	10	9	90	2	20	8	80	3	37	5	6
40	1	10	9	90	3	30	7	70	1	13	7	8
45	3	30	7	70	3	30	7	70	1	13	7	8
50	2	20	8	80	2	20	8	80	3	37	5	6
55	2	20	8	80	3	30	7	70	2	25	6	7
60	2	20	8	80	4	40	6	60	1	13	7	8
65	2	20	8	80	4	40	6	60	2	25	6	7
70	1	10	9	90	4	40	6	60	2	25	6	7:
75	1	10	9	90	4	40	6	60	3	37	5	6
80	1	10	9	90	4	40	6	60	3	37	5	6
85	1	10	9	90	3	30	7	70	1	13	7	8
90	2	20	8	80	4	40	6	60	1	13	7	8
95	2	20	8	80	3	30	7	70	1	13	7	8
100	3	30	7	70	3	30	7	70	1	13	7	8
105	3	30	7	70	4	40	6	60	2	25	6	7
110	3	30	7	70	3	30	7	70	2	25	6	7
115	3	30	7	70	3	30	7	70	2	25	6	7
120	3	30	7	70	3	30	7	70	2	25	6	7
Average	2	16	8	84	3	30	7	70	2	29	6	7

Table 1 Number and percentage of larvae in light and dark area







(Sawin-McCormack *et al.*, 1995; Rodriguez and Campos, 2009) and becomes photophilic with age (Godoy-Herrera *et al.*, 1984) or adult-hood (Gong *et al.*, 2010). Such transition may be due to the development from immersed foraging stage to active wandering stage in search of place for pupation (Humberg and Sprecher, 2017; Bainbridge and Bownes, 1981). In contrast, Yamanaka *et al.* (2013) reported that wandering larvae are photo-negative and prefer to pupate in darkness. Furthermore, von Essen *et al.* (2011) made an interesting observation that larva prefer food in darkness over food in light. Besides, Warrick *et al.* (1999) reported that photo-negativity of the larva can be elicited by light with wave length ranging from UV to green. Additionally, studies found that, changes in light intensity elicit turning behavior (Hassan *et al.*, 2000; Scantlebury *et al.*, 2007).

Further, to know the relevance of this experiment/ study to the school students, it was demonstrated to the pre-service teachers (of RIE, Mysuru) and in-service teachers (of JNVs). They opined that this can be an effective project for the higher secondary students to train them in scientific method and to go beyond text book.

#### CONCLUSION

It may be concluded from the above results that, all the three strains (O.K, se and wh) of larvae are negative phototaxic, and among them normal O.K larvae are more sensitive to light than the two mutants. There may exist a relation between phototaxis response and eye color. Further, as the pre and in-service teachers opined, this experiment can be an effective project for the higher secondary students for active learning by doing and to go beyond text book.

#### Acknowledgement

Authors sincerely thank Mr. Santhosh, Physics section, I/C Zoology section, Head DESM and the Principal of RIE,

Mysuru and the Director, NCERT, New Delhi, for their moral support and encouragement.

#### References

- Bainbridge SP, Bownes M (1981). Staging the metamorphosis of *Drosophila melanogaster*. J Embryol Exp Morphol, 66: 57-80.
- Ball SB, Catherine CE and Christian R (2006). Technology improves learning in large principles of economics classes: using our WITS. *Am Econ Rev Pap Proced*, 96(2): 442-446.
- CarlFriedrich RW, Thomas RC (2017). *Drosophila* connectomics: mapping the larval eye's mind. *Current Biol*, 27(21); R1161-R1163.
- Emerson T and Taylor B (2004). Comparing student achievement across experimental and lecture-oriented sections of a principle of microeconomics course. *Southern Econ J*, 703: 672-693.
- Emery P, Stanewsky R, Helfrich-Förster C, Emery-Le M, Hall JC, Rosbash, M (2000). *Drosophila* CRY is a deep brain circadian photoreceptor. *Neuron*, 26: 493-504.
- Farca Luna AJ, von Essen AM, Widmer YF, Sprecher SG (2013). Light preference assay to study innate and circadian regulated photobehavior in *Drosophila* larvae. *J Vis Exp*, 20(74).
- Frank B (1997). The impact of classroom experiments on the learning of economics: an empirical investigation. *Econ Inquiry*, 35(4):763.
- Gershow M, Berck M, Mathew D, Luo L, Kane E A, Carlson J R, *et al.* (2012). Controlling airborne cues to study small animal navigation. *Nat Methods*, 9: 290-296.
- Godoy-Herrera R, Burnet B, Connolly K, Gogarty J (1984). The development of locomotor-activity in *Drosophila melanogaster* larvae. Heredity, 52: 63-75.
- Gong Z (2009). Behavioral dissection of *Drosophila* larval phototaxis. *Biochem Biophys Res Commun*, 382(2): 395-399.
- Gong Z, Gong Z (2012). A molecular diffusion based utility model for *Drosophila* larval phototaxis. *Theor Biol Med Model*, 2(9): 3.
- Gong Z, Liu J, Guo C, Zhou Y, Teng Y, Liu L (2010). Two pairs of neurons in the central brain control *Drosophila* innate light preference. *Sci*, 22; 330(6003): 499-502.
- Hassan J, Busto M, Iyengar B, Campos AR (2000). Behavioral characterization and genetic analysis of the *Drosophila melanogaster* larval response to light as revealed by a novel individual assay. *Behav Genet*, 30, 59-69.
- Hassan J, Iyengar B, Scantlebury N, Rodriguez Moncalvo V, Campos A R (2005). Photic input pathways that mediate the *Drosophila* larval response to light and circadian rhythmicity are developmentally related but functionally distinct. *J Comp Neurol*, 481: 266-275.
- Hassan J, Iyengar B, Scantlebury N, Rodriguez Moncalvo V, Campos AR (2005). Photic input pathways that mediate the *Drosophila* larval response to light and circadian rhythmicity are developmentally related but functionally distinct. *J Comp Neurol*, 17; 481(3): 266-275.
- Hernandez-Nunez L, Belina J, Klein M, Si G, Claus L, Carlson JR, Samuel AD (2015). Reverse-correlation

analysis of navigation dynamics in *Drosophila* larva using optogenetics. *Elife*, 5; 4.

- Humberg T-H and Sprecher SG (2017). Age- and wavelength-dependency of *Drosophila* larval phototaxis and behavioral responses to natural lighting conditions. *Front Behav Neurosci*, 11:66.
- Kane EA, Gershow M, Afonso B, Larderet I, Klein M, Carter A R, et al. (2013). Sensorimotor structure of *Drosophila* larva phototaxis. *Proc Natl Acad Sci U S A*, 110: E3868-E3877.
- Kane EA, Gershow M, Afonso B, Larderet I, Klein M, Carter AR, de Bivort BL, Sprecher SG, Samuel AD (2013).
  Sensorimotor structure of *Drosophila* larva phototaxis. *Proc Natl Acad Sci U S A*, 110(40): E3868-3877.
- Keene AC, Mazzoni EO, Zhen J, Younger MA, Yamaguchi S, Blau J, Desplan C, Sprecher SG (2011). Distinct visual pathways mediate *Drosophila* larval light avoidance and circadian clock entrainment. J *Neurosci*, 27; 31(17): 6527-34.
- Malpel S, Klarsfeld A, Rouyer F (2002). Larval optic nerve and adult extra-retinal photoreceptors sequentially associate with clock neurons during *Drosophila* brain development. *Dev*, 129(6): 1443-1453.
- Mazzoni EO, Desplan C, Blau J (2005). Circadian pacemaker neurons transmit and modulate visual information to control a rapid behavioral response. *Neuron*, 45: 293-300.
- Mishra AK, Tsachaki M, Rister J, Ng J, Celik A, Sprecher SG (2013). Binary cell fate decisions and fate transformation in the *Drosophila* larval eye. *PLoS Genet*, 9(12): e1004027.
- Nagaraj G (2016). Comparison of phototaxis responses using improvised apparatus: A novel experiment for hands-on and minds-on learning. *Global J Mul Dis Stud*, 5(9): 82-91.
- NCERT (2006). Position paper 1.1: National focus group on teaching of Science, 11-17.
- Rodriguez MVG, Campos AR (2009). Role of serotonergic neurons in the *Drosophila* larval response to light. BMC *Neurosci*, 23(10): 66.
- Salcedo E, Huber A, Henrich S, Chadwell L V, Chou, W H, Paulsen, R. *et al.* (1999). Blue- and green-absorbing visual pigments of Drosophila: ectopic expression and physiological characterization of the R8 photoreceptor cell-specific Rh5 and Rh6 rhodopsins. *J Neurosci*, 19: 10716-10726.
- Sawin-McCormack E P, Sokolowski MB, Campos AR (1995). Characterization and genetic analysis of *Drosophila melanogaster* photobehavior during larval development. *J Neurogenet*, 10: 119-135.
- Sawin-McCormack EP, Sokolowski MB, Campos AR (1995). Characterization and genetic analysis of *Drosophila melanogaster* photobehavior during larval development. *J Neurogenet*, 10(2): 119-135.
- Scantlebury N, Sajic R, Campos A R (2007). Kinematic analysis of *Drosophila* larval locomotion in response to intermittent light pulses. *Behav. Genet*, 37: 513-524.
- Sprecher S G, Pichaud F, Desplan C (2007). Adult and larval photoreceptors use different mechanisms to specify the same Rhodopsin fates. *Genes Dev.* 21: 2182-2195.

- Sprecher SG, Pichaud F, Desplan C (2007). Adult and larval photoreceptors use different mechanisms to specify the same Rhodopsin fates. *Genes Dev*, 21(17): 2182-2195.
- Stanewsky R, Kaneko M, Emery P, Beretta B, Wager-Smith K, Kay S A, Rosbash M, Hall JC (1998). The cryb mutation identifies cryptochrome as a circadian photoreceptor in Drosophila. *Cell*, 95: 681-692.
- von Essen AM, Pauls D, Thum AS, Sprecher SG (2011). Capacity of visual classical conditioning in *Drosophila* larvae. *Behav Neurosci*, 125: 921-929.
- Warrick J M, Vakil M F, Tompkins L (1999). Spectral sensitivity of wildtype and mutant *Drosophila melanogaster* larvae. *J Neurogenet*, 13: 145-156.
- Xiang Y, Yuan Q, Vogt N, Looger LL, Jan LY, Jan YN (2010). Light-avoidance-mediating photoreceptors tile the *Drosophila* larval body wall. *Nature*, 468(7326): 921-926.
- Yamanaka N, Romero N M, Martin FA, Rewitz KF, Sun M, O'Connor M. B, et al. (2013). Neuroendocrine control of *Drosophila* larval light preference. Sci, 341: 1113-1116.
- Zhao W, Gong C, Ouyang Z, Wang P, Wang J, Zhou P, Zheng N, Gong Z (2017). Turns with multiple and single head cast mediate *Drosophila* larval light avoidance. *PLoS One*, 12(7): e0181193.

### How to cite this article:

Nagaraj G.2018, Do Drosophila Larvae Prefer Light or Dark?-an Improvised Investigatory Project For High School Students To Go Beyond Text Book. *Int J Recent Sci Res.* 9(1), pp. 23023-23027. DOI: http://dx.doi.org/10.24327/ijrsr.2018.0901.1383

\*\*\*\*\*\*