



ISSN: 0976-3031

Available Online at <http://www.recentscientific.com>

CODEN: IJRSFP (USA)

International Journal of Recent Scientific Research
Vol. 10, Issue, 04(E), pp. 31924-31927, April, 2019

**International Journal of
Recent Scientific
Research**

DOI: 10.24327/IJRSR

Research Article

EVALUATION OF BIOCHEMICAL RESPONSES OF ONION (*ALLIUM CEPA* L.) SEEDLINGS UNDER DROUGHT STRESS

Vidya vani M¹, Osman basha P² and Riazunnisa K*¹

¹Department of Biotechnology and Bioinformatics, Yogi Vemana University, Kadapa, Andhra Pradesh, India

²Department of Genetics and Genomics, Yogi Vemana University, Kadapa, Andhra Pradesh, India

DOI: <http://dx.doi.org/10.24327/ijrsr.2019.1004.3364>

ARTICLE INFO

Article History:

Received 4th January, 2019

Received in revised form 25th

February, 2019

Accepted 23rd March, 2019

Published online 28th April, 2019

Key Words:

Drought stress, Onion, Proline, Phenols,
Relative water content.

ABSTRACT

Drought is one of the most important global problems for agriculture. Plants adapt and respond to water stress by inducing different defense mechanisms by changing their cellular metabolism. The present study is aimed to gauge the effect of drought (with holding watering for six days) by screening the nine cultivars of onion seedlings, by monitoring the following parameters like chlorophyll content, relative water content (RWC), quantitative analysis of proteins, sugars, proline, phenols, flavonoids, and tannins. Drought stress decreased the chlorophyll content in all the cultivars. Relative water content was maintained in Arka lalima under stress when compared with other cultivars. Protein, sugars, and proline content increased in all cultivars under drought conditions. Maximum proline levels were examined in prema 178, arka kirthaman and arka lalima under stress conditions. Secondary metabolites such as flavonoids, phenols and tannins levels enhanced in stressed seedlings. Among all nine cultivars of onion arka kirthaman, arka lalima and prema 178 showed highest response to drought stress when compared to the control. Further physiological and molecular investigation is needed to know the drought tolerance mechanism in onion cultivars.

Copyright © Vidya vani M, Osman basha P and Riazunnisa K, 2019, this is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

The scarcity of fresh water for agriculture around the world threatens the productivity of crop lands. Irregular distribution or scarcity of most of the rainfall in rain fed areas at any specific stage of the crop leads to drought (Bouman and Aureus, 2009). Four-tenths of the world's agricultural land lies in arid and/or semi-arid regions. Water stress is one of the most serious environmental stresses which limit production by regulating plant growth and development. Crop plants habituate to such drought stress conditions by invoking various physiological, biochemical and molecular responses (Maksup *et al.* 2014). The change in response include the over production of enzymes responsible for the biosynthesis of detoxification enzymes and osmolytes (Huseynova *et al.* 2016). Habash *et al.* 2014 reported physiological and a transcriptome response in durum wheat under drought stress. Glycine betaine, proline, sugars and some inorganic ions are small compatible solutes synthesized and accumulated under water stress (Chaves *et al.*, 2003). These solutes maintain cellular dehydration and membrane integrity and this in turn furnish tolerance against cellular dehydration and drought (Ramanjulu and Bartels, 2002). Sucrose, stachyose, sorbitol, raffinose,

galactinol and trehalose are sugars included in raffinose family oligosaccharides. These sugars play important role in the drought tolerance mechanism of plants and seeds (Seki *et al.*, 2007). In addition to all these, reactive oxygen species is also synthesized in drought stress.

Proline, an amino acid play a versatile role in the water stress defense mechanisms. Non-enzymatic scavenging mechanisms include accumulation of proline, glycine betaine (Ashraf *et al.*, 2007). Amino acid proline is deposited in big amount in response to environmental stresses and it occurs widely in higher plants (Ozturk *et al.*, 2002), proline acts as an compatible solute for osmotic adjustment. In plants secondary metabolites are known to increase during water stress. Accumulation of secondary metabolites observed by Bartwal *et al.* 2013 is a defense mechanism of plants to adapt themselves under drought stress by altering their cellular metabolism. Increased synthesis of flavonoid content in wheat leaves under water stress was observed by Ma *et al.* 2014.

*Corresponding author: Riazunnisa K

Department of Biotechnology and Bioinformatics, Yogi Vemana University, Kadapa, Andhra Pradesh, India

MATERIALS AND METHODS

Onion seeds Collection, Germination and Drought Treatment

Nine cultivars of onion seeds were selected and collected (Agrifound rose, Bellary, Nasik red, Prema-178, Arka Kirthiman, Arka Lalima, Gavran Light Red, and Light Red & Red Diamond) and allowed to germinate for 10 days at 27 °C in plant growth chamber. Seasons and places where they are cultivated are reported by Sai Sudha *et al.* 2019. The 10 day old seedlings were subjected to non lethal drought stress by withholding watering for six days under controlled growth chamber conditions.

Chlorophyll and Relative water content (RWC)

Onion seedlings (0.05g) were homogenized in 80% acetone (v/v) for estimating the total chlorophyll content (Arnon 1949). For relative water content, 20 seedlings from each cultivar were weighed immediately (FW) after harvesting (Sai Sudha *et al.* 2015). Relative water content was calculated by the following formula.

$$\text{Relative water content} = \frac{\text{FW}-\text{DW}}{\text{TW}-\text{DW}} \times 100$$

Where, FW- Fresh weight, DW- Dry weight, TW- Turgid weight.

Proteins, Sugars and Proline levels

Determination of protein concentrations was done by Lowry method (Lowry *et al.* 1951). The total soluble carbohydrates extraction was made with 50 mg of seedlings and sugars were determined by anthrone method. Proline was assayed according to the method described by Bates *et al.* (1973). Fresh seedlings of 500 mg were homogenized in 5 ml of 3 % (w/v) sulphosalicylic acid. Acid ninhydrin and glacial acetic acid were added and incubated for 1 h in a boiling water bath at 90°C followed by an ice bath then toluene was added and measured at 560 nm against blank.

Total Phenols, Total Flavonoid and Tannin Content

The total phenolic content was determined by the spectrophotometer method (Harborne 1973). 1 ml of sample was mixed with Folin- Ciocalteu phenol reagent and Na₂CO₃ and mixed thoroughly. The mixture was kept in the dark for 90 min at 23°C, and then the absorbance was read at 750 nm. The TPC was expressed as milligrams of Gallic acid equivalents (GAE) per g of dried sample. The total flavonoid content of the sample was determined by using ammonium chloride colorimetric method as described by Chang *et al.* (2002) with slight modifications. Absorbance of the mixture was measured at 415 nm and the calibration curve was prepared by using quercetin as standard. Tannins were quantified according to Van-Burden and Robinson (1981). 0.5 g of the sample was mixed with distilled water in a mechanical shaker. To the Filtrate 0.1 M FeCl₃ and 0.1 N HCl and 0.008 M potassium ferrocyanide was added and absorbance was read at 120 nm.

Statistical analysis

RESULTS AND DISCUSSION

Chlorophyll Content and Relative water Content

Total chlorophyll content reduced in all cultivars under drought stress (Fig. 1). Negative correlation between chlorophyll

content and drought has been reported many researchers which is in accordance with our results (Lakshmi sahitya *et al.* 2018). When compared to controls relative water content varied and decreased in all the cultivars under drought. In controls relative water content level was maximum in Light red i.e. 91 % and minimum in Nasik red i.e. 72 % (Fig. 1). Under stress conditions more relative water content was observed in Arka lalima and lesser quantity in Gavran light red.

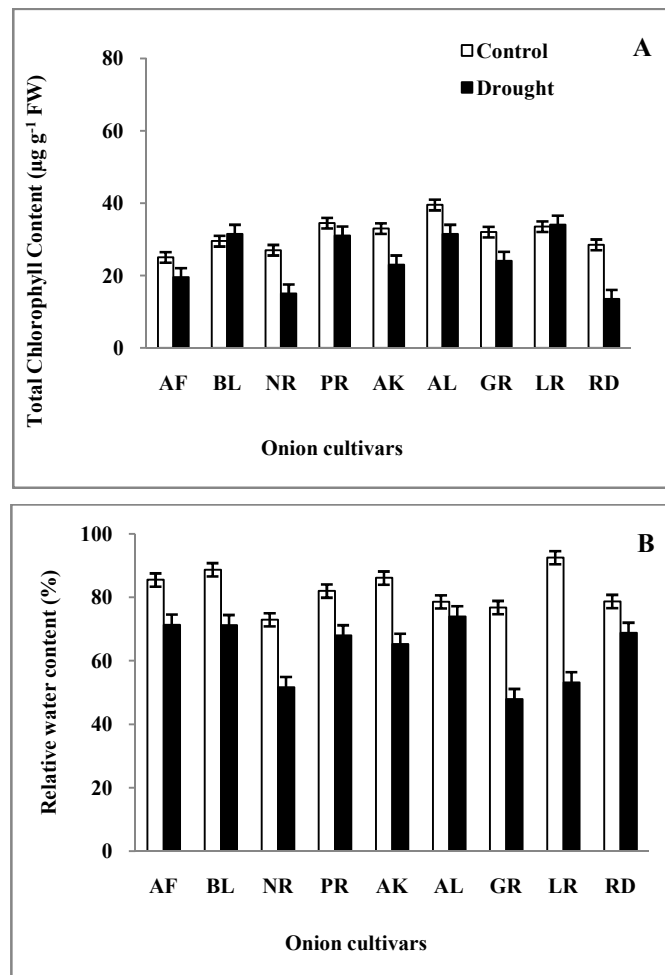


Figure 1 Effect of drought stress on (A) total chlorophyll content and (B) relative water content in different onion seedlings. Each bar represents the mean (\pm SE) of three replications.

Protein, Sugar and Proline levels

Under withholding water condition protein, sugars and proline accumulation varied and increased in all the cultivars when compared with controls (Fig. 2). Overexpression of drought-inducible galactinol synthase gene (*ArGOLS2*) in transgenic *Arabidopsis* enhanced drought tolerance because of the accumulation of galactinol and raffinose (Taji *et al.*, 2002). Maximum protein quantity was observed in Arka kirthiman and Prema-178 and minimum in Nasik red (Fig. 2). Highest proline content was monitored in Arka kirthiman and Arka lalima in stress. The correlation between the accumulation of proline and tolerance of drought stress has been demonstrated by overexpression of proline-5-carboxylate synthase gene *P5CS* or by antisense suppression of the proline dehydrogenase (*ProDH*) gene in various plants (Bartels and Ramanjulu 2005).

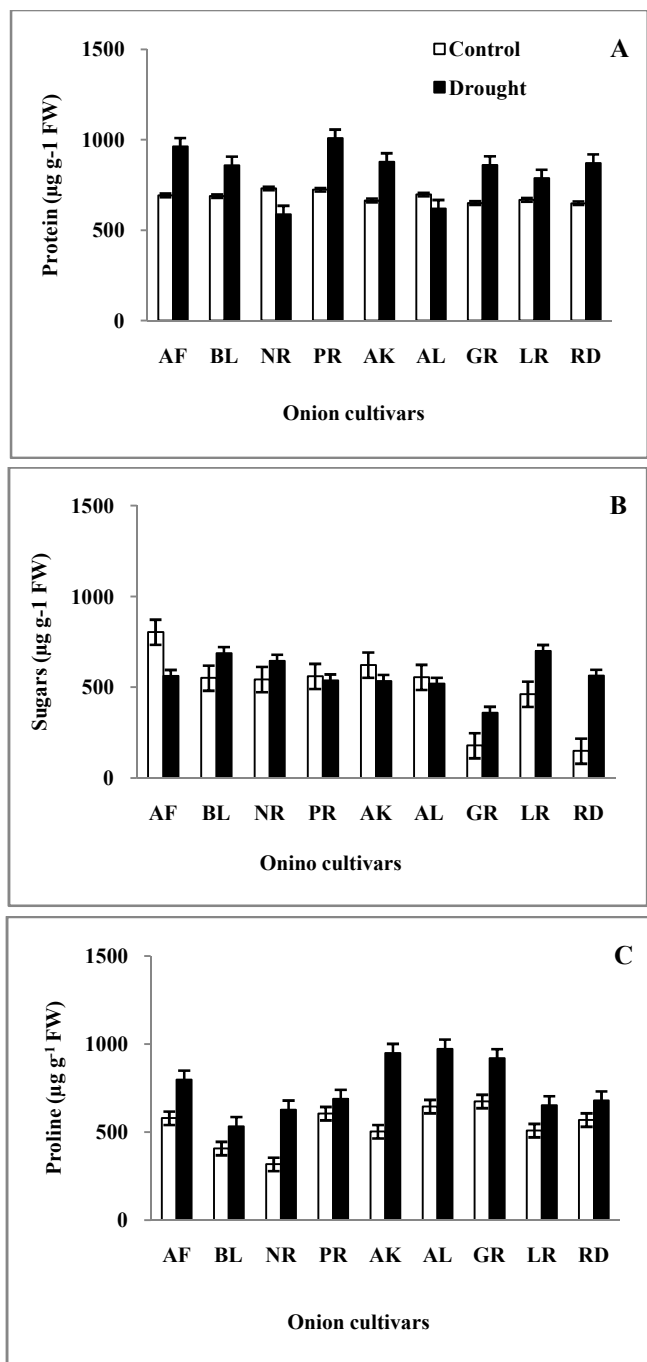


Figure 2 Effect of drought stress on (A) Protein (B) Sugars and (C) Proline in different onion seedlings. Each bar represents the mean (\pm SE) of three replications.

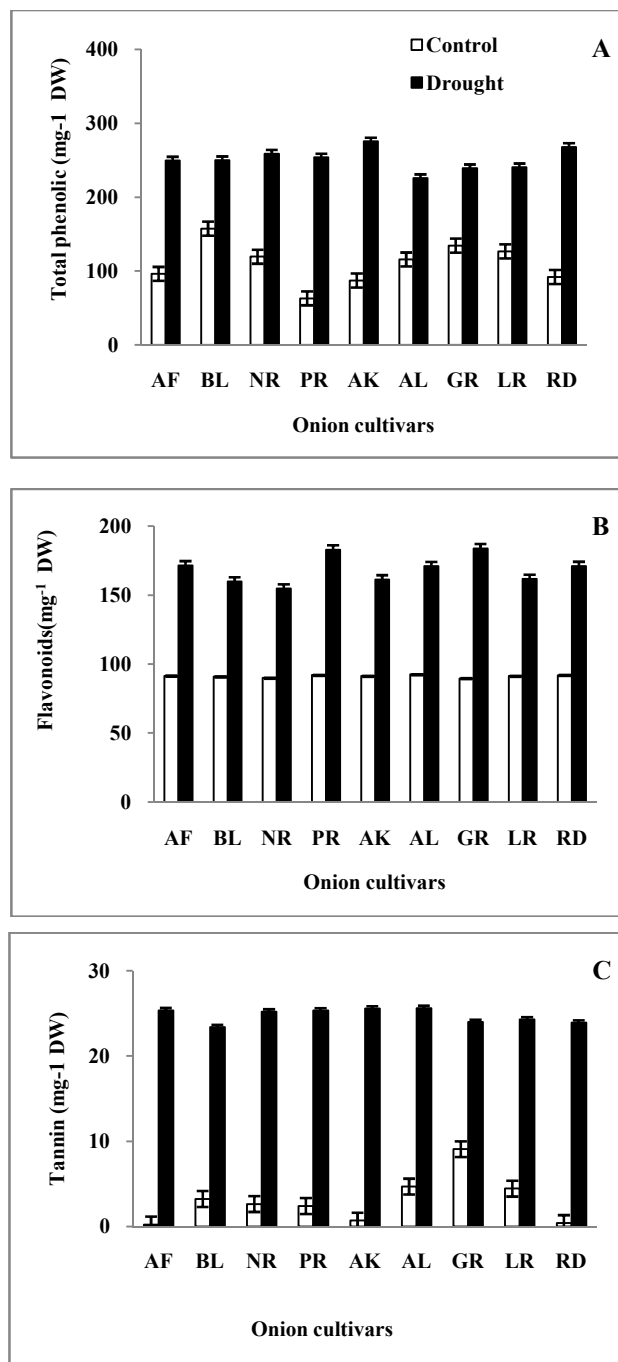


Figure 2 Effect of drought stress on (A) Phenol (B) flavonoids and (C) tannins in different onion seedlings. Each bar represents the mean (\pm SE) of three replications.

Total phenols, Total flavonoid and Tannin content

Drought stress enhanced total phenols, flavonoids content in amaranthus leafy vegetable (Sarker and Oba 2018). Total phenol, flavonoid and tannin content of the nine cultivars were different both in control and drought (Fig. 3). Greatest total phenol, flavonoid and tannin levels were observed in all the cultivars under drought conditions. More phenolic content were examined in Arka kirthiman and Prema-178. Under drought stress highest flavonoid levels were observed in Arka kirthiman, Prema-178 and Gavran light red (Fig. 3). The increase in tannin content was very high in stress when compared with controls in all the cultivars.

CONCLUSION

The present investigation was performed to evaluate the physiological and biochemical responses of onion germplasm under withheld water stress. Assessment of Nine onion cultivars revealed the negative correlation between chlorophyll, relative water content, and positive correlation between proteins, sugars, proline, phenols, flavonoids, tannins and drought. Three onion germplasm like arka kirthiman, arka lalima and prema 178 exhibited better response among all the cultivars.

Acknowledgements

This research work was financially supported to Dr. K.R from Department of Science and Technology (No.SR/FT/LS-352/2012).

Reference

1. Arnon, D. I. (1949). Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant physiology*, 24(1), 1.
2. Lowry, O. H., Rosebrough, N. J., Farr, A. L., & Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *J Biol Chem*, 193(1), 265-275.
3. Bates, L. S., Waldren, R. P., & Teare, I. D. (1973). Rapid determination of free proline for water-stress studies. *Plant and soil*, 39(1), 205-207.
4. Harborne, J. B. (1973). Phenolic compounds. In *Phytochemical methods* (pp. 33-88). Springer Netherlands.
5. Chang, C. C., Yang, M. H., Wen, H. M., & Chern, J. C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of food and drug analysis*, 10(3).
6. Van-Burden, T. P., & Robinson, T. (1981). The biochemistry of alkaloids.
7. Taji T, Ohsumi C, Luchi S, Seki M, Kasuga M (2002): Important roles of drought and cold inducible genes for galactinol synthase in stress tolerance in *Arabidopsis thaliana*. *The Plant Journal* 29: 417-426.
8. Bartels D, Ramanjulu S (2005): Drought and stress tolerance in plants. *Critical Reviews in Plant Sciences* 24: 23-58.
9. Maksup S, Roytrakul S, Supaibulwatana K (2014) Physiological and comparative proteomic analyses of Thai jasmine rice and two chick cultivars in response to drought stress. *J Plant Interact* 9:43-55
10. Habash DZ, Baudo M, Hindle M, Powers SJ, Defoin-Platel M, et al. (2014) Systems Responses to Progressive Water Stress in Durum Wheat. *PLoS ONE* 9(9):108431. doi:10.1371/journal.pone.0108431
11. U. Lakshmi Sahitya, M. S. R. Krishna, R. Sri Deepthi, G. Shiva Prasad and D. Pedd Kasi (2018) Seed Antioxidants Interplay with Drought Stress Tolerance Indices in Chilli (*Capsicum annuum* L) Seedlings. *Biomed research international* <https://doi.org/10.1155/2018/1605096>
12. Irada M. Huseynova, Samira M. Rustamova, • Saftar Y. Suleymanov, Durna R. Aliyeva, Alamdar Ch. Mammadov, Jalal A. Aliyev (2016) Drought-induced changes in photosynthetic apparatus and antioxidant components of wheat (*Triticum durum* Desf.) varieties. *Photosynth Res*. DOI 10.1007/s11120-016-0244-z
13. Bouman, B and Aureus, A. 2009. Every drop counts. *Rice Today*. IRRI Publishers, pp 16-18.
14. Chaves, M.M., J.P.Maroco, J.pereira, 2003.under standing plants response to drought from genes to the whole plant. *Functional plant Biol*; 30,239-264
15. Ramanjulu, S and Bartels D. 2002. Drought and desiccation- induced modulation of gene expression in plants. *Plant Cell Environ*, 25(2)-141-151
16. Seki M, Umezawa T, Urano K, Shinozaki K (2007): Regulatory metabolic networks in drought stress responses. *Current Opinion in Plant Biology* 10:296-302.
17. Ozturk L, Demir Y (2002): In vivo and in vitro protective role of proline. *Plant Growth Regulation* 38:259-264.
18. Ashraf M, Foolad M R (2007) Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany* 59:206-216.
19. Sarker U and Oba S (2018) Drought stress enhances nutritional and bioactive compounds, phenolic acids and antioxidant capacity of *Amaranthus leafy vegetable* Sarker and Oba *BMC Plant Biology* 18:258 <https://doi.org/10.1186/s12870-018-1484-1>
20. Ma D, Sun D, Wang C, Li Y, Guo T (2014) Expression of flavonoid biosynthesis genes and accumulation of flavonoid in wheat leaves in response to drought stress. *Plant physiology and Biochemistry*. 80: 60-66
21. Bartwal A, Mall R, Lohani P, Guru SK, Arora S (2013) Role of secondary metabolites and brassinosteroids in plant defense against environmental stresses. *Journal of plant growth regulation*. 32 (1) 216-232.
22. G. Sai Sudha, P. Ramesh, A. Chandra Sekhar, T. Sai Krishna, P.V. Brahmachari, K. Riazunnisa Genetic diversity analysis of selected Onion (*Allium cepa* L.) germplasm using specific RAPD and ISSR polymorphism markers. *Biocatalysis and Agricultural Biotechnology* 2019 17 : 110-118.
23. G. Sai Sudha and K. Riazunnisa Effect of salt stress (NaCl) on morphological parameters of onion (*Allium cepa* L.) seedlings *International Journal of Plant, Animal and environmental sciences* 2015 Vol 5(4): 125-128.

How to cite this article:

Vidya vani M, Osman basha P and Riazunnisa K., 2019, Evaluation of Biochemical Responses of Onion (*Allium Cepa* L.) Seedlings Under Drought Stress. *Int J Recent Sci Res*. 10(04), pp. 31924-31927. DOI: <http://dx.doi.org/10.24327/ijrsr.2019.1004.3364>
